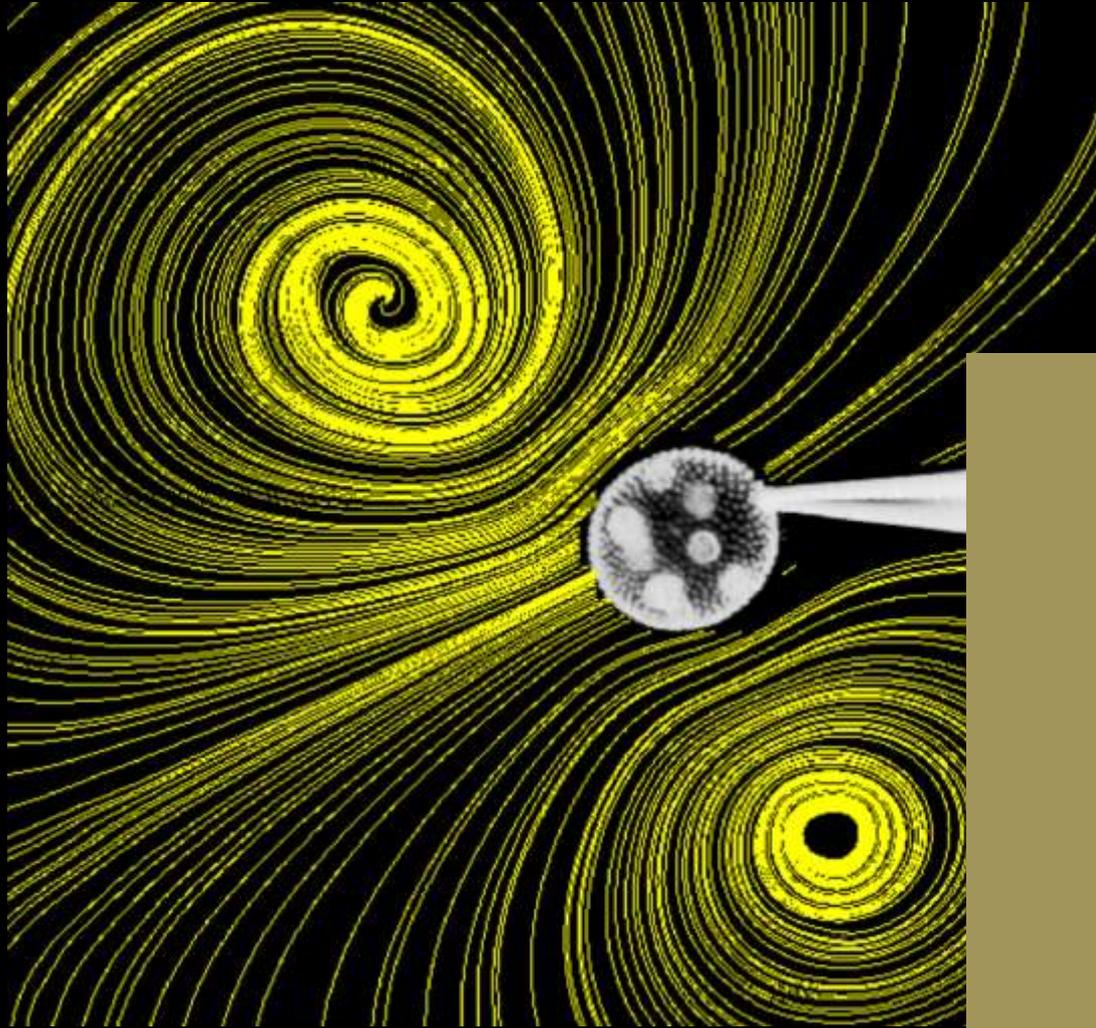


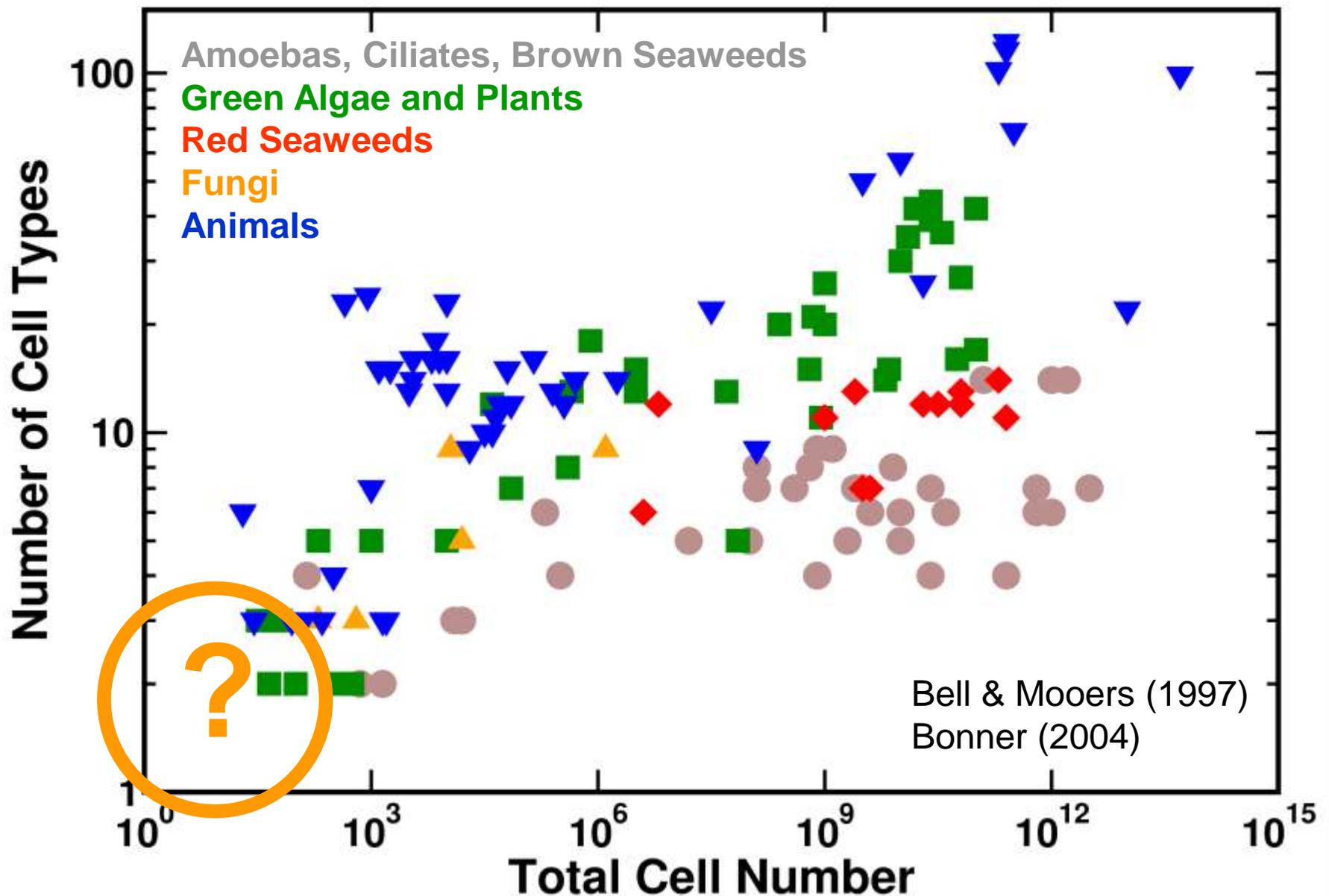
Stirring Tails of Evolution

R E Goldstein
DAMTP



www.damtp.cam.ac.uk/user/gold
www.youtube.com/Goldsteinlab

The Size-Complexity Relation



The Recent Literature

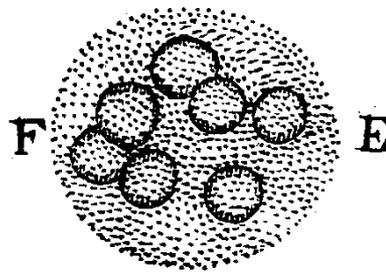
IV. Part of a Letter from Mr Antony Van Leeuwenhoek, concerning the Worms in Sheeps Livers, Gnats, and Animalcula in the Excrements of Frogs.

When I brought these particles before the Magnifying-glass, I did not only see that they were round, but that the outward skin of them was quite set over with many protuberant parts, which did seem to me to be triangular, and pointed towards the end; so that it seemed to me, that in the great circle of the roundness, stood such particles, all orderly and equally from each other; so that on a small body did stand about two thousand of the before-mentioned convex or protuberant particles.

This was to me a very pleasant sight, because the said particles, as often as I did look on them, did never lye still, and that their motion did proceed from their turning round; and that the more, because I did fancy at first that they were small animals, and the smaller these particles were, the greener was their colour; and on the contrary, in the greatest, that were as big as a great corn of sand, there was no green colour at all to be discerned on the outside.

These particles had each of them within included 5, 6, 7, nay, some to 12 small round globules, of the same shape as the body was wherein they were included.

Fig: 5



Phil. Trans. 22,
509-518 (1700)

CAROLI LINNÆI

EQUITIS DE STELLA POLARI,
ARCHIATRI REGII, MED. & BOTAN. PROFESS. UPSAL.;
ACAD. UPSAL. HOLMENS. PETROPOL. BEROL. IMPER.
LOND. MONSPEL. TOLOS. FLORENT. SOC.

SYSTEMA NATURÆ

(1758)

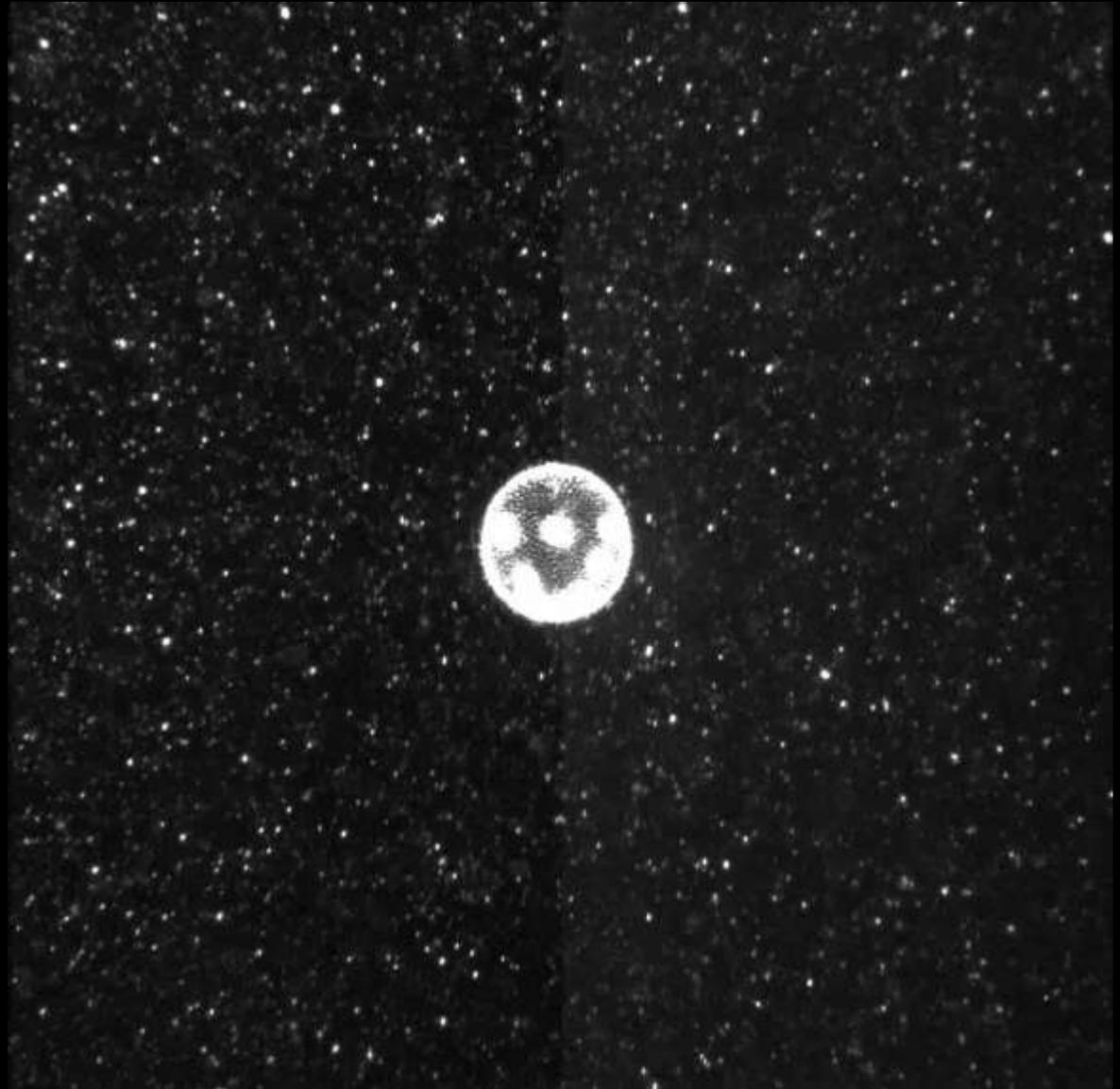
312. VOLVOX. *Corpus liberum, gelatinosum, rotundatum, artubus destitutum.*

Proles subrotundi, nidulantes, sarsi.

Volvendo seque rotando celeriter movens absque artubus! viviparus natis, nepotibus, pronepotibus, abnepotibus conspicuis intra animalculum minutissimum.

Volvox In Its Own Frame

Tracking microscope
in vertical orientation
Laser sheet illumination
of microspheres

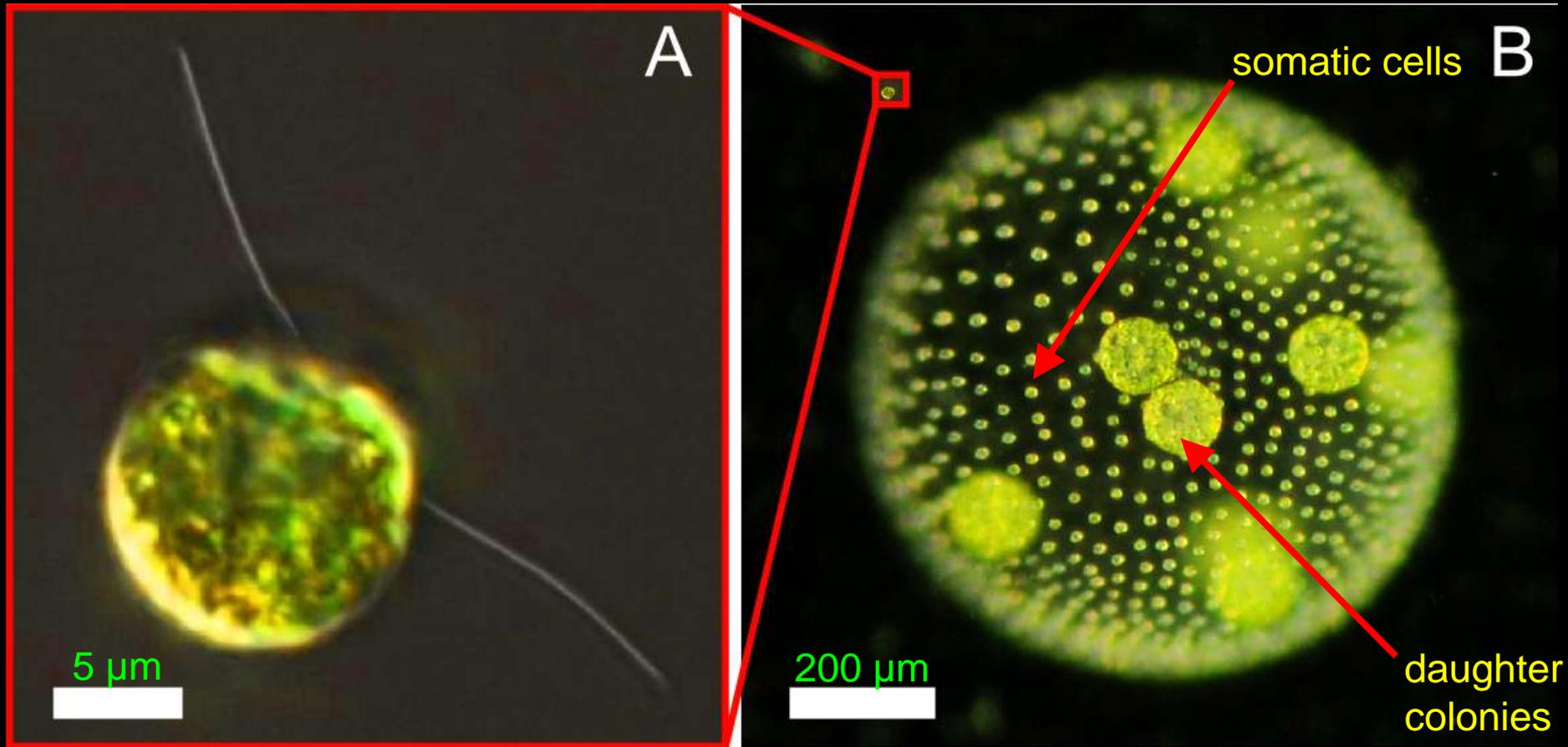


Drescher, Goldstein, Michel, Polin, and Tuval, *preprint* (2010)

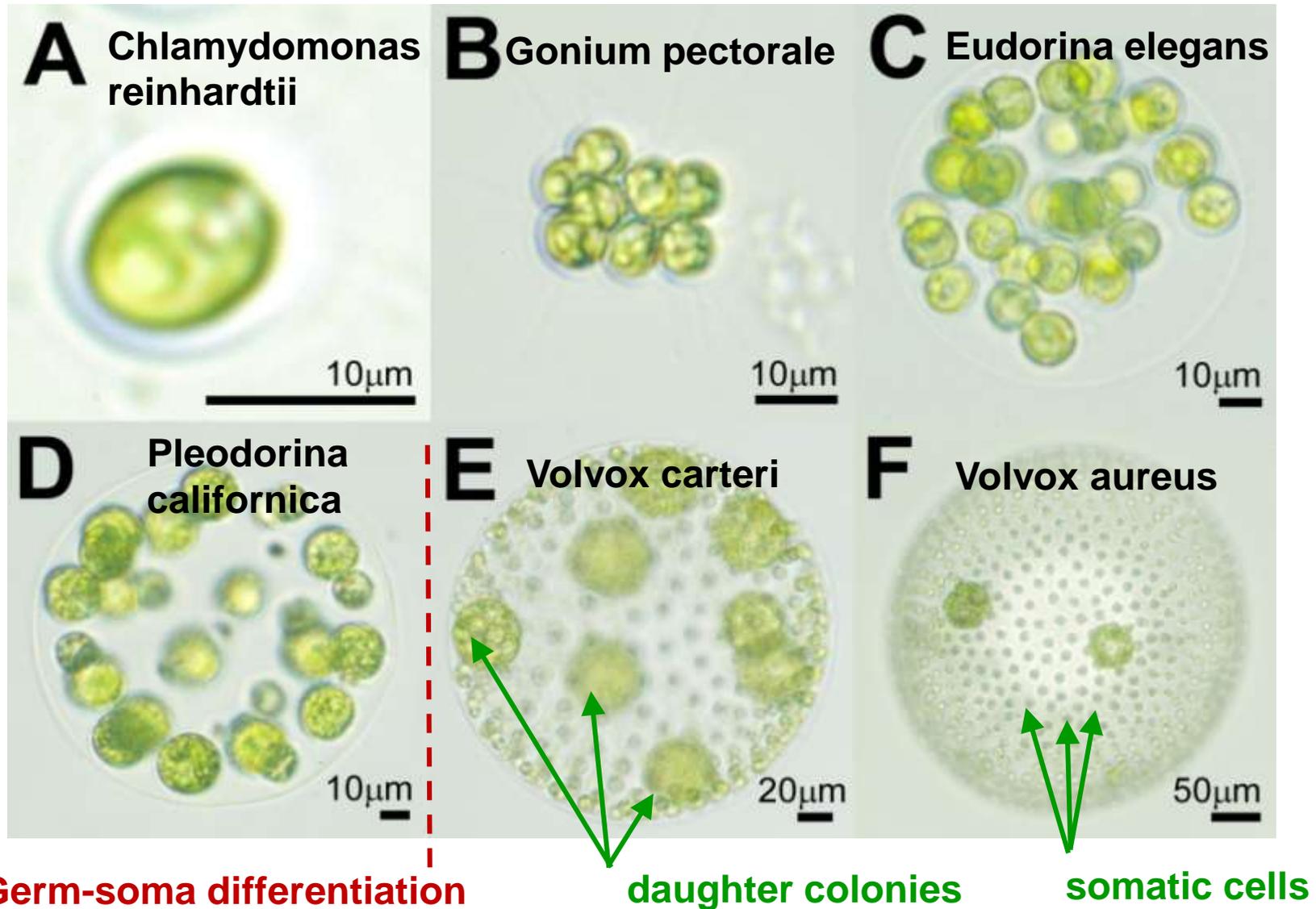
Evolutionary Transition to Multicellularity

Chlamydomonas reinhardtii

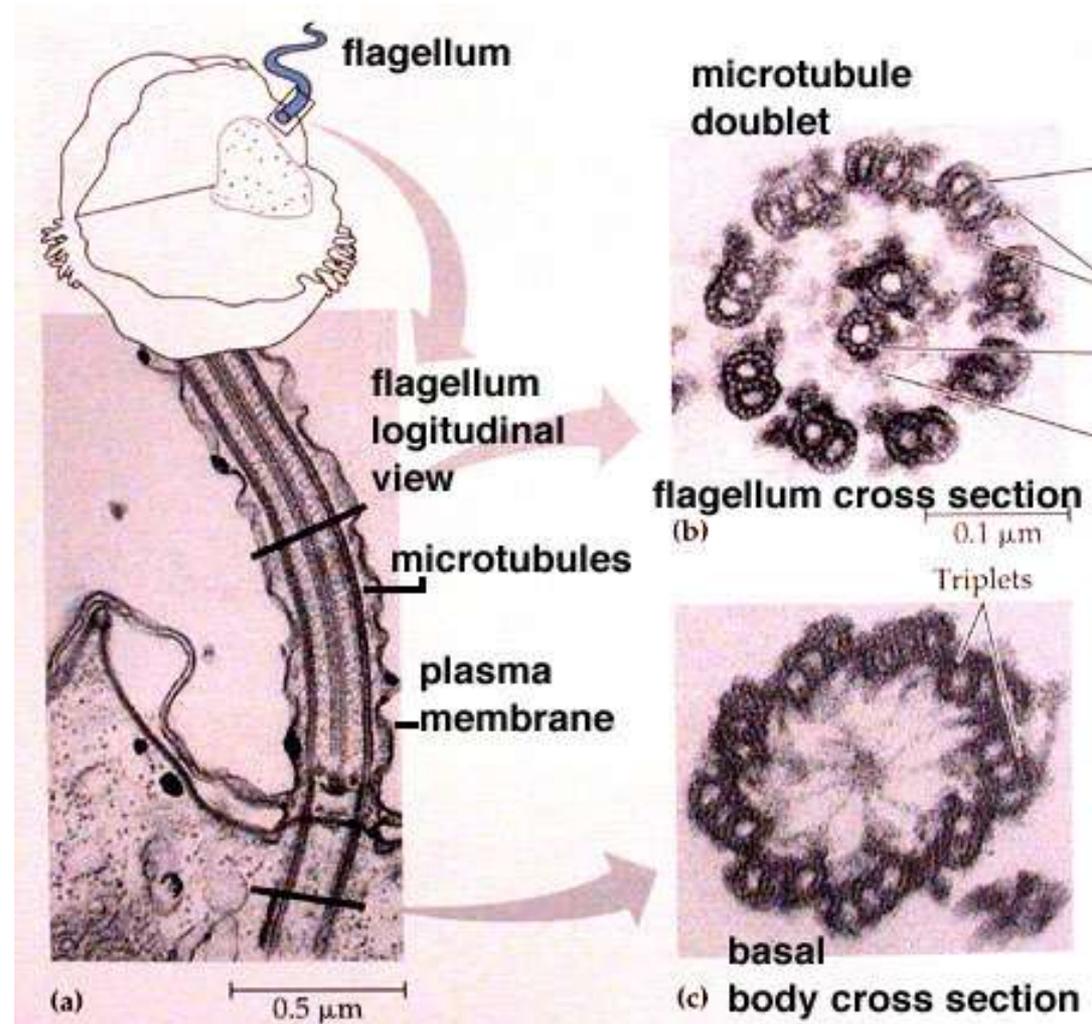
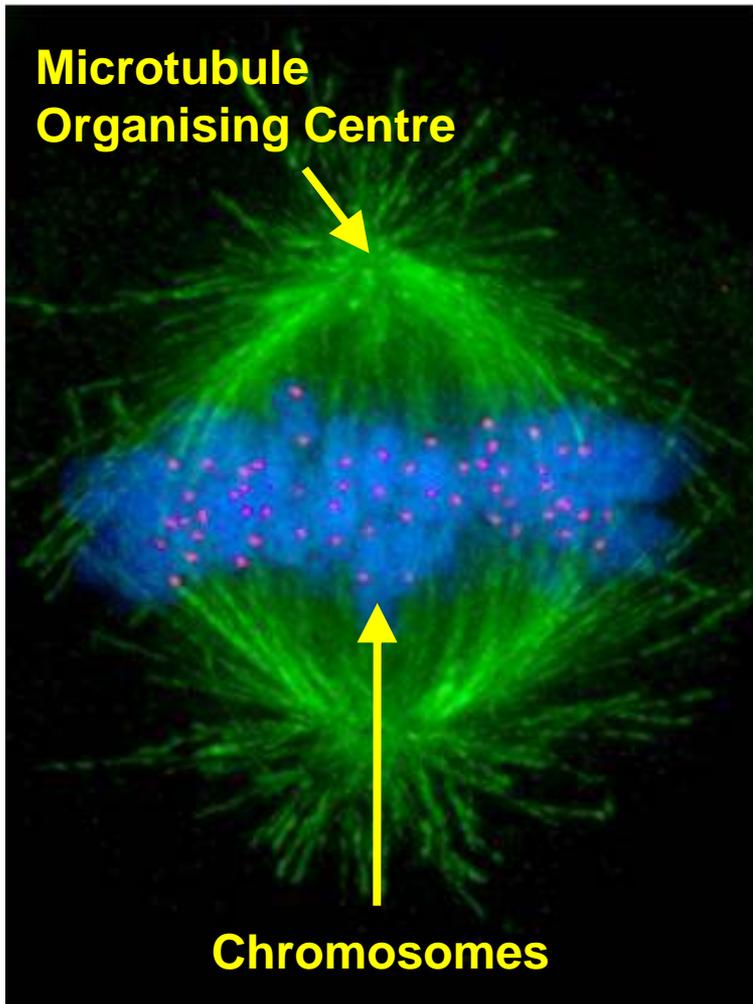
Volvox carteri



A Family Portrait



Structure of Flagella & the Flagellation Constraint



Basal bodies are microtubule organizing centres
...flagella are resorbed during cell division (no multi-tasking)
(Bell & Koufopanou, '85,'93)

Advection & Diffusion

If a fluid has a typical velocity \mathbf{U} , varying on a length scale L , with a molecular species of diffusion constant \mathbf{D} . Then there are two times:

We define the Péclet number as the ratio:

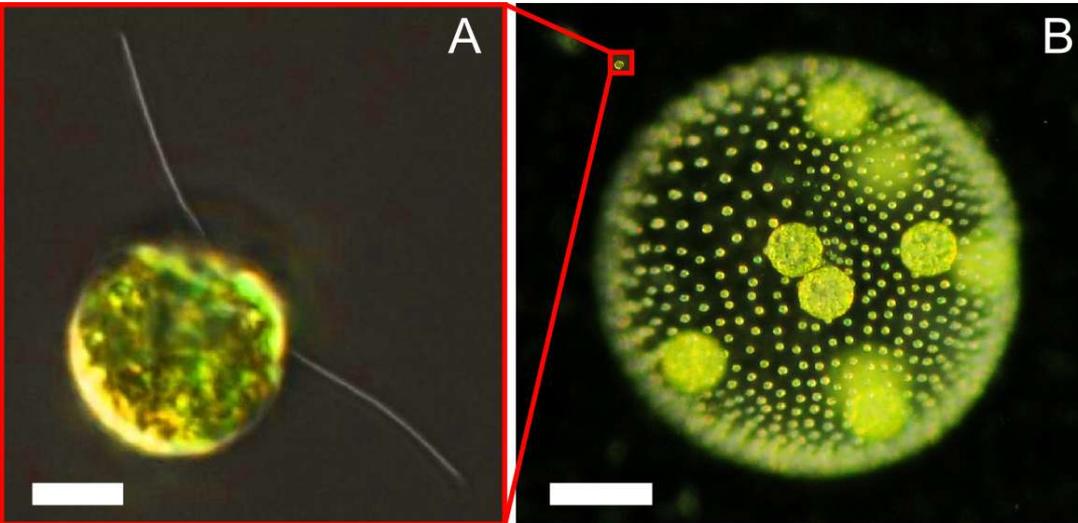
$$Pe = \frac{t_{diffusion}}{t_{advection}} = \frac{UL}{D}$$

This is like the Reynolds number comparing inertia to viscous dissipation:

$$Re = \frac{UL}{\nu}$$

$$t_{advection} = \frac{L}{U}$$

$$t_{diffusion} = \frac{L^2}{D}$$



If $U=10 \mu\text{m/s}$, $L=10 \mu\text{m}$,
 $Re \sim 10^{-4}$, $Pe \sim 10^{-1}$

*At the scale of an individual cell,
diffusion dominates advection.*

*The opposite holds for
multicellularity...*

Solari, Ganguly, Michod, Kessler, Goldstein, *PNAS* (2006)

Short, Solari, Ganguly, Powers, Kessler & Goldstein, *PNAS* (2006)

Life Cycles of the Green and Famous



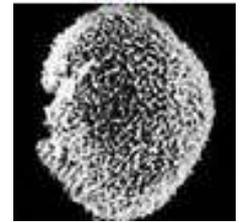
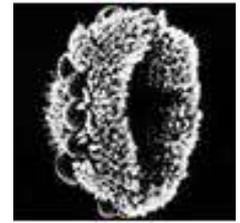
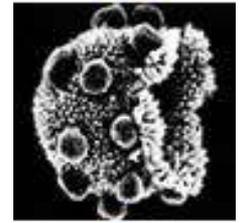
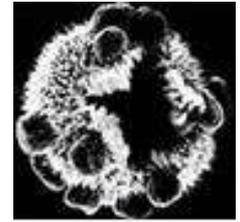
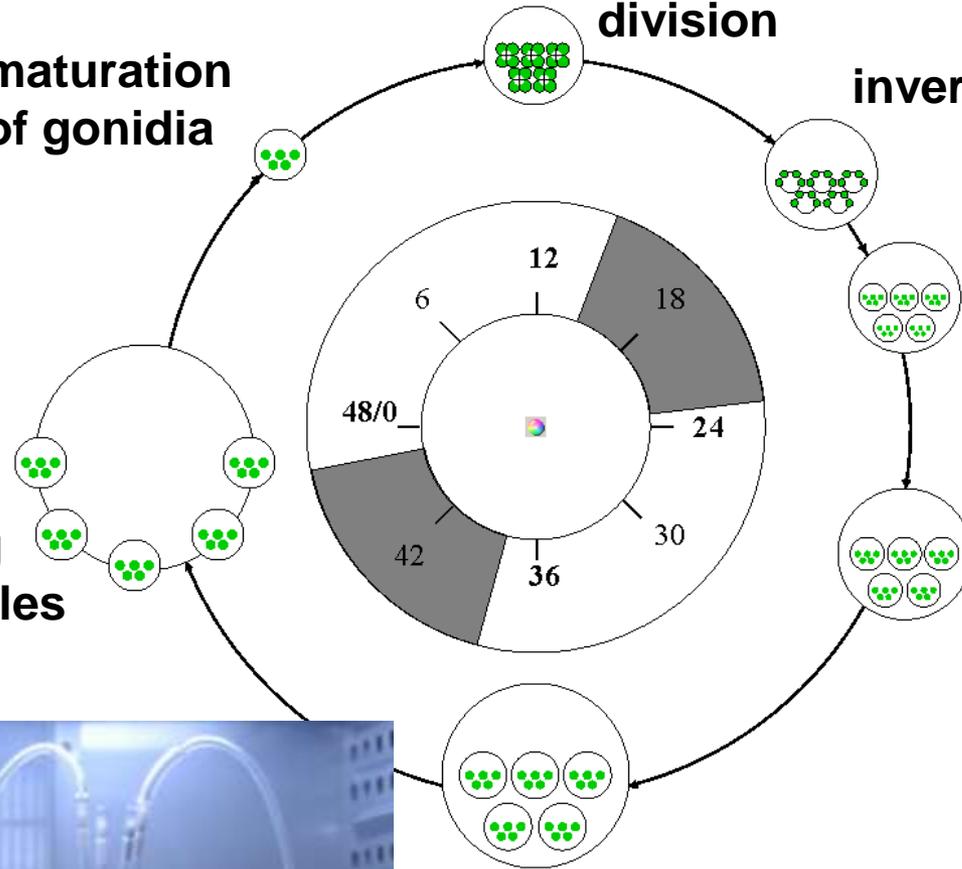
maturation
of gonidia

division

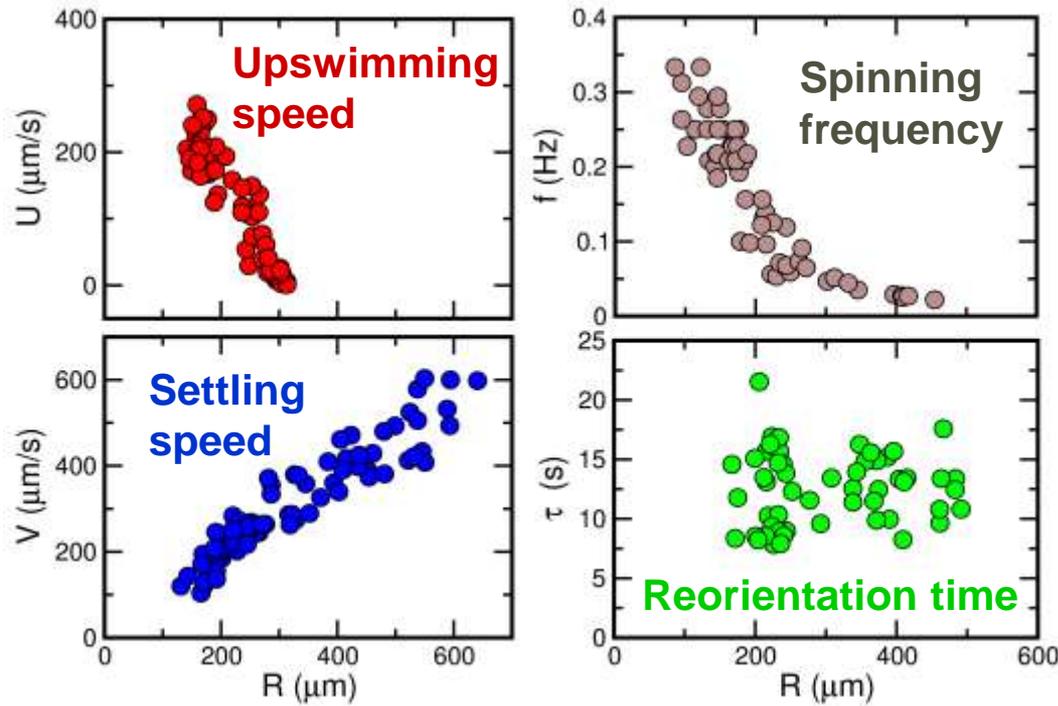
inversion

hatching
of juveniles

cytodifferentiation
and expansion



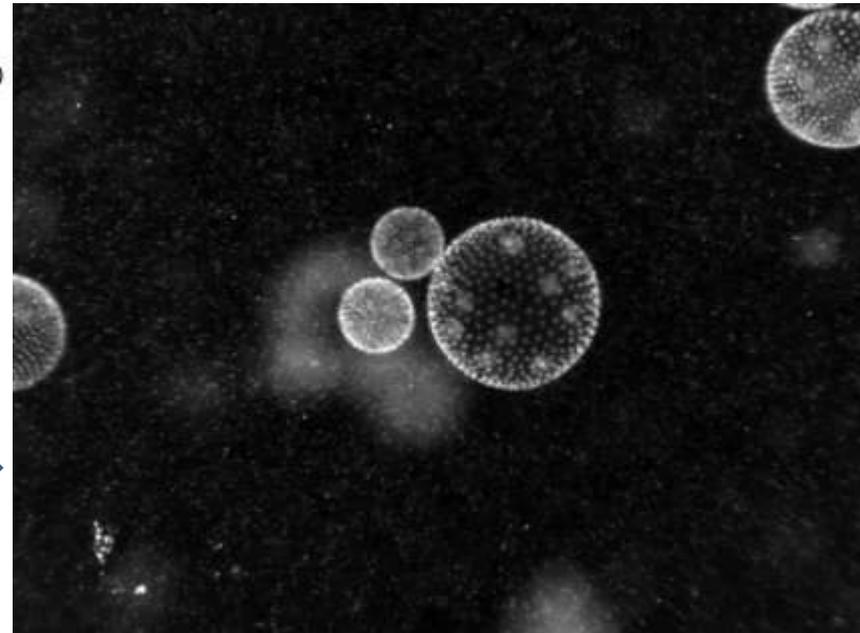
Volvox as a Model Organism for Biological Physics



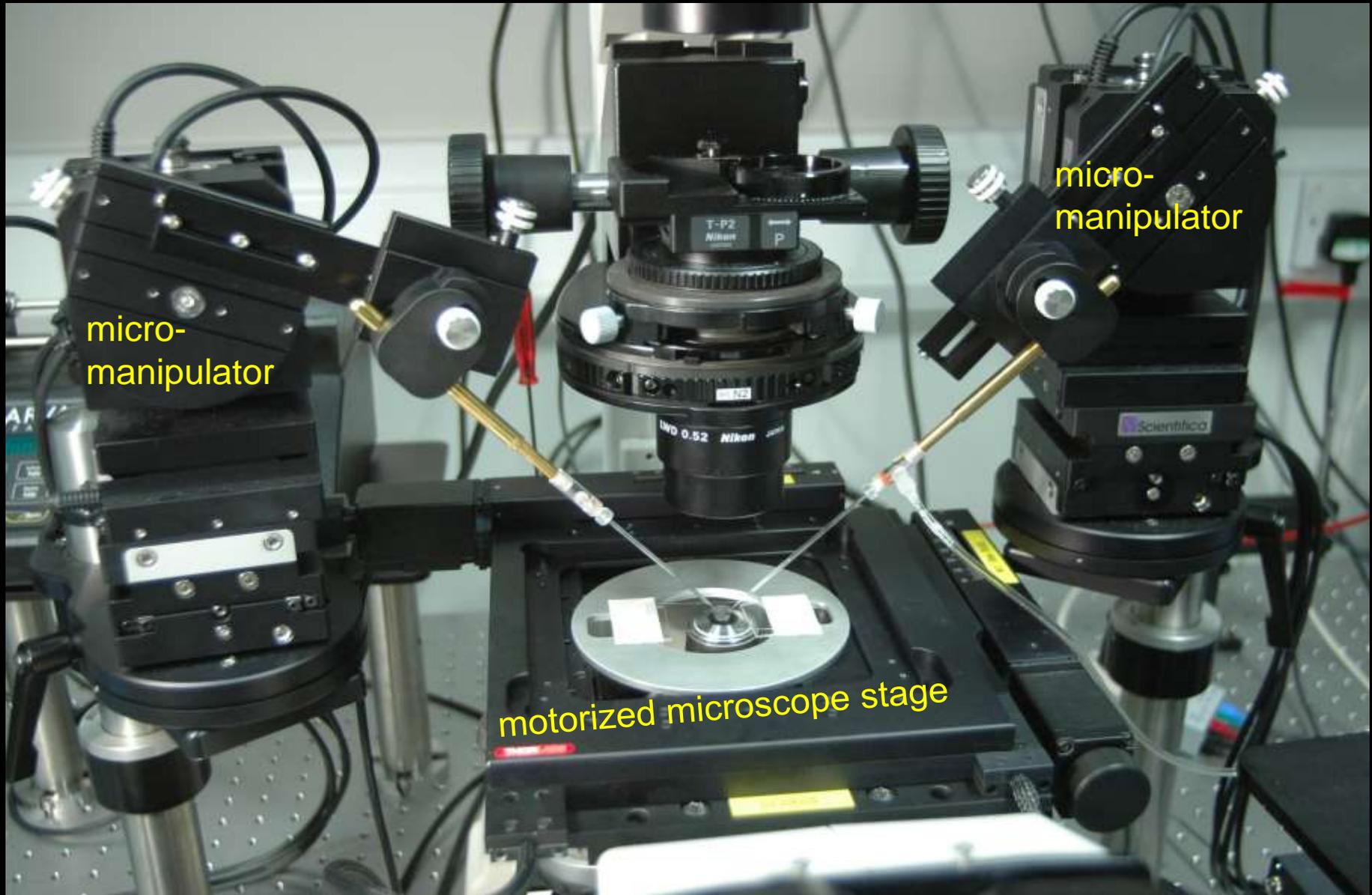
As the colonies grow they move more systematically through the parameter space of swimming, sinking, spinning, and self-righting.



As an example, Volvox can exhibit “hydrodynamic bound states” which can be explained quantitatively using mutually advected force singularities.



Microscopy & Micromanipulation



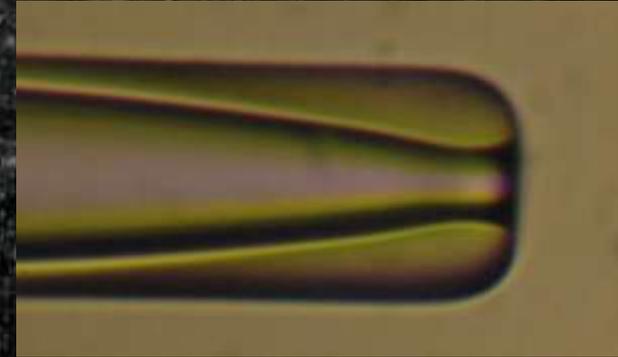
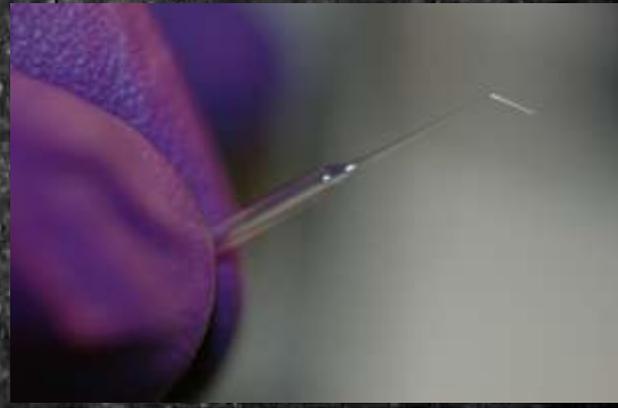
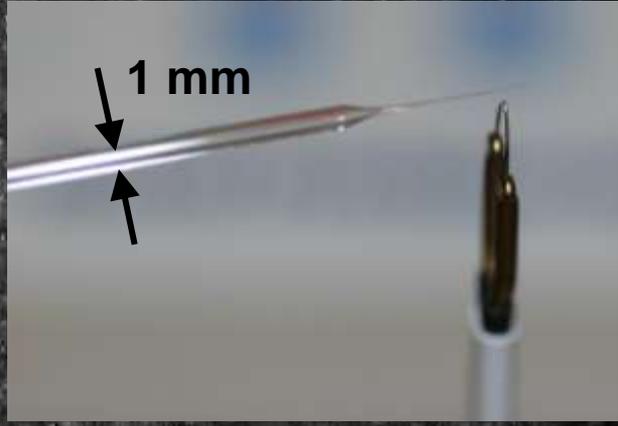
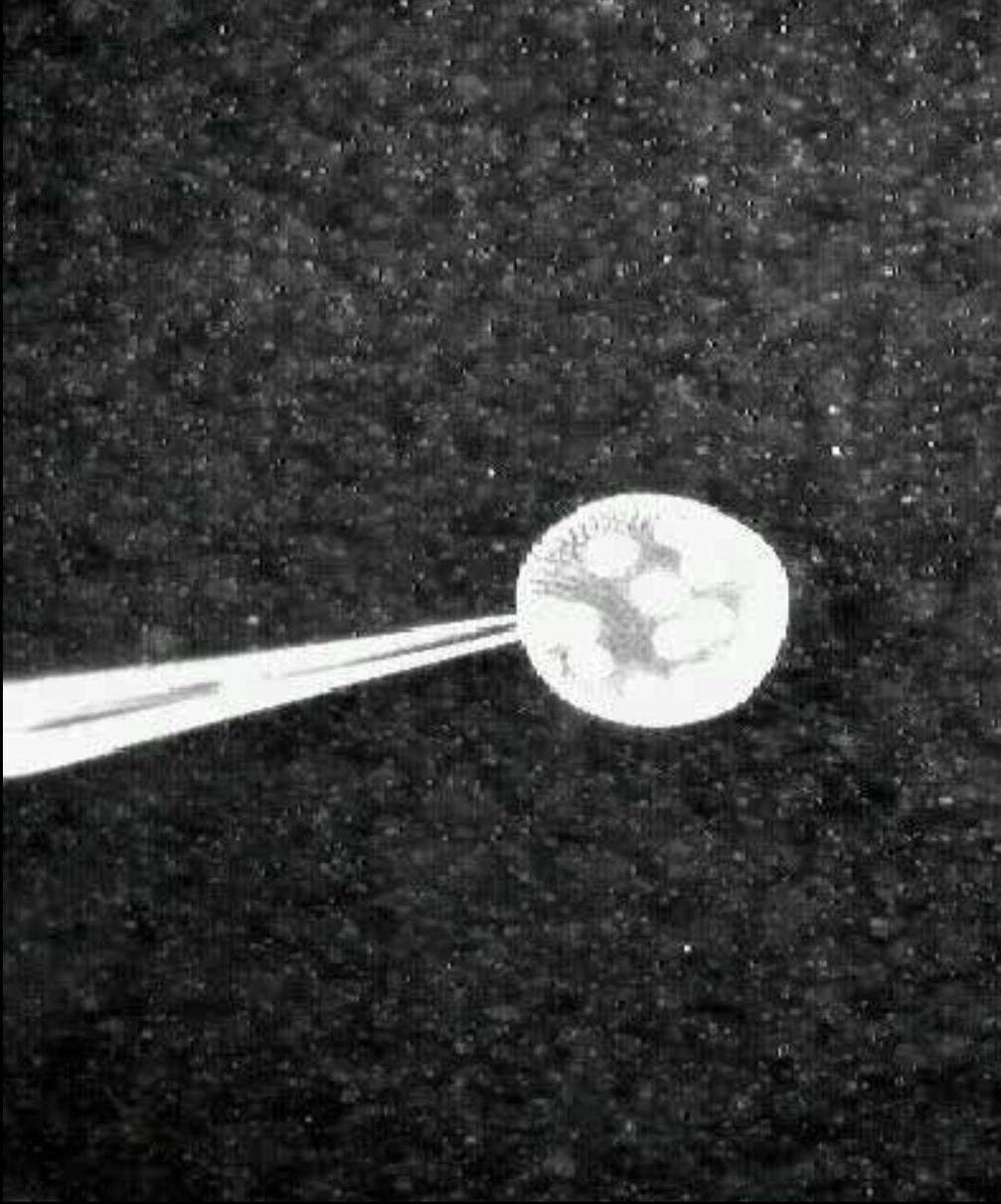
micro-manipulator

micro-manipulator

motorized microscope stage

Stirring by *Volvox carteri*

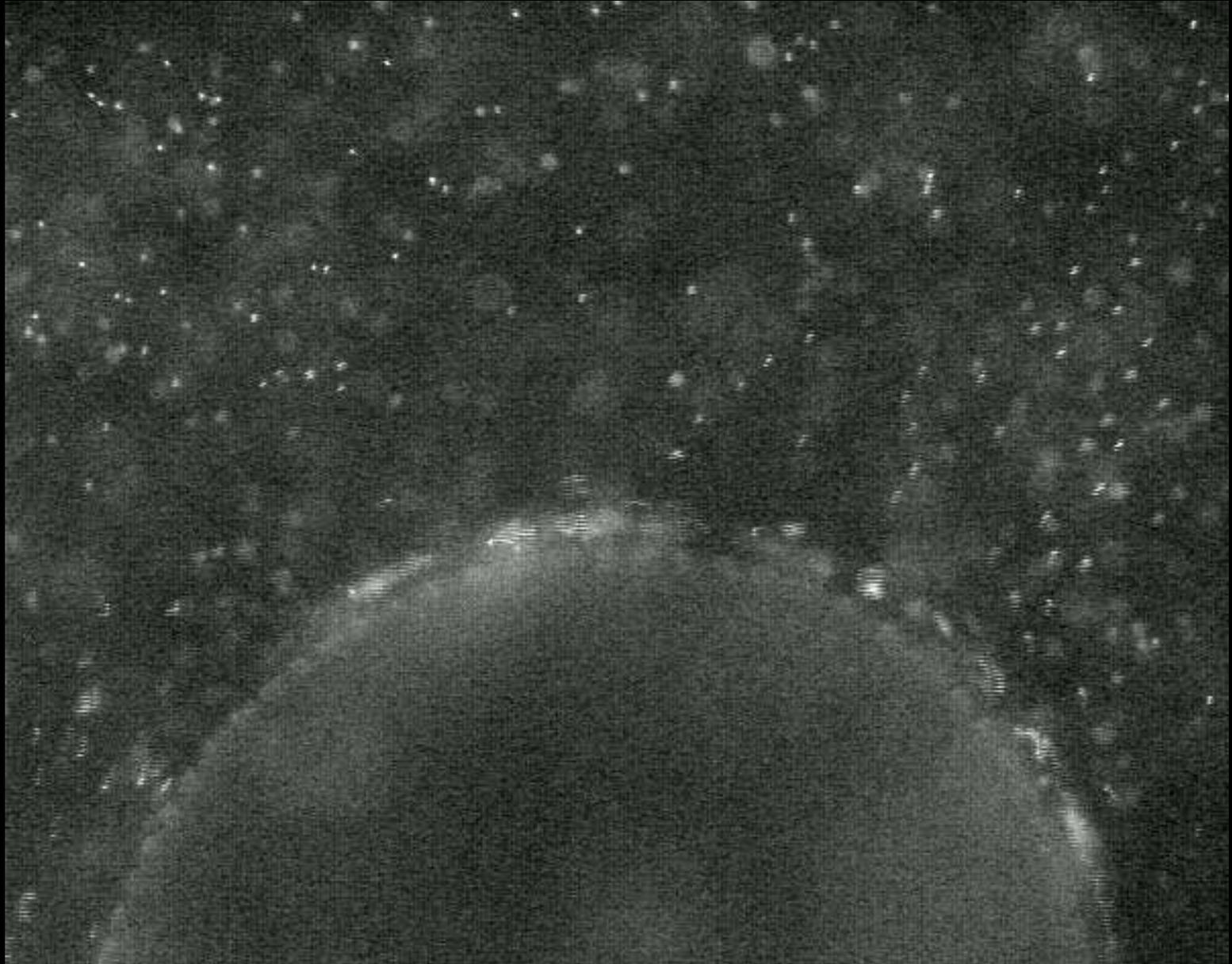
Pseudo-darkfield (4x objective, Ph4 ring)



Tools of the trade – micropipette preparation

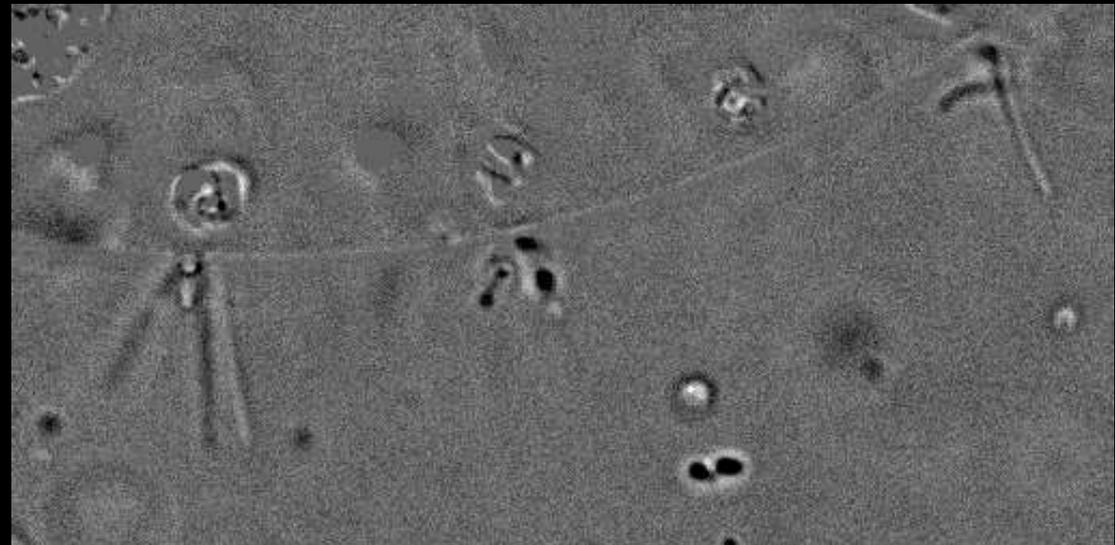
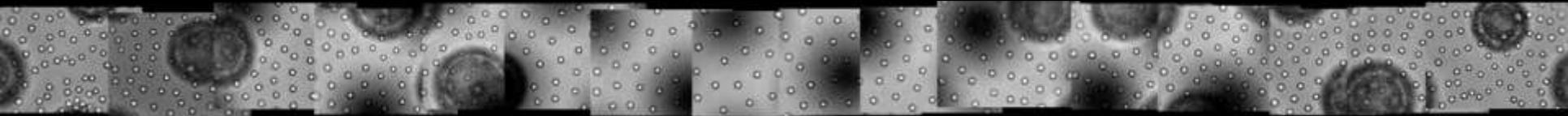
A Closer View

Fluorescence



Flagella Beating/Symmetry

(2000 frames/s
background
subtraction)



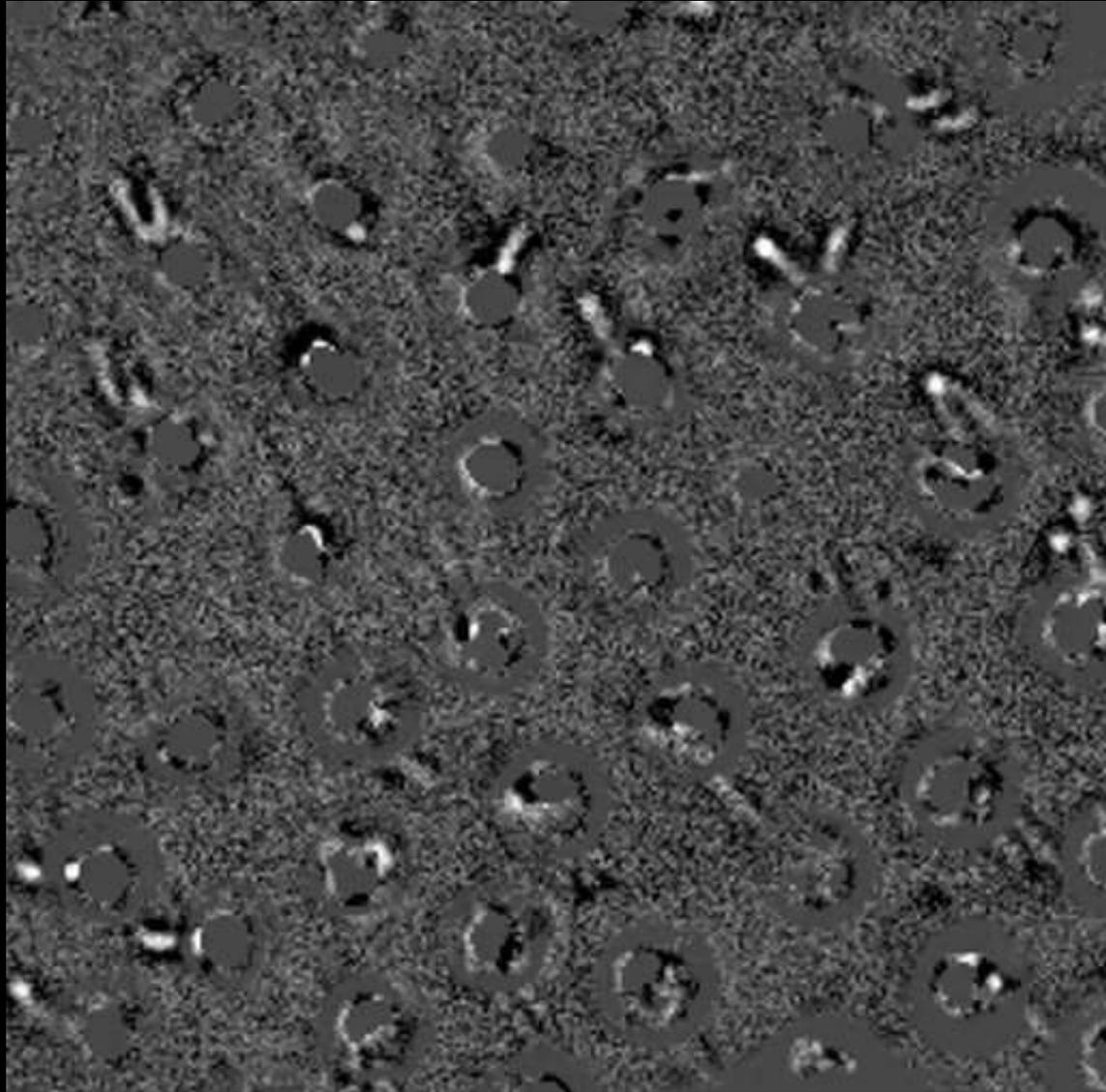
Polin, Tuval,
Drescher
& Goldstein (2009)

Metachronal Waves in *Volvox* (Side View)



Polin, Tuval, Pesci, Goldstein (2010)

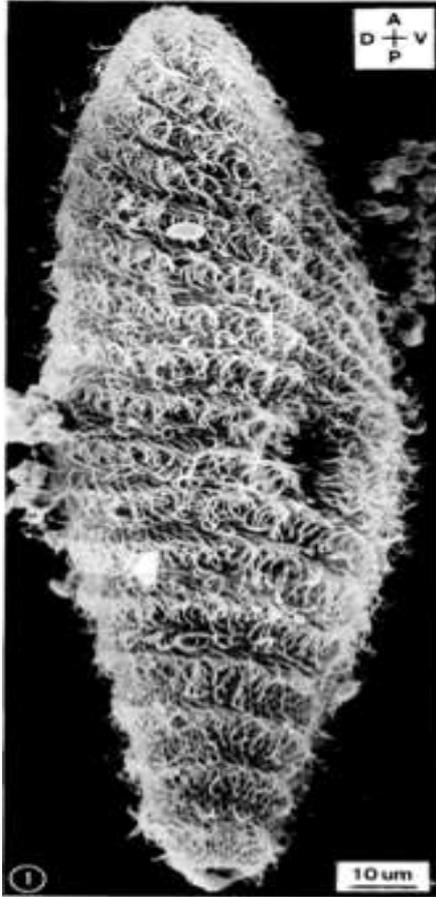
Metachronal Waves in *Volvox* (Top View)



Polin, Tuval, Pesci, Goldstein (2010)

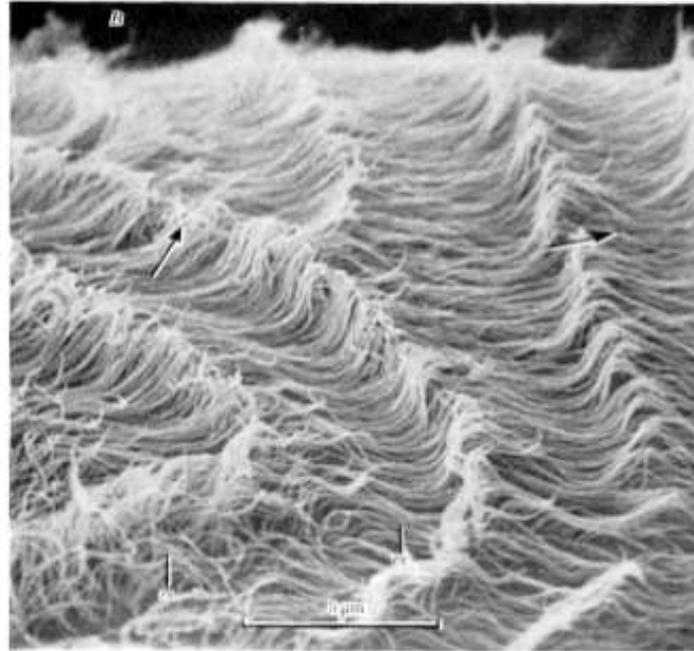
Flagellar Coordination in Eukaryotes

Paramecium
(protozoan)



30 μm

Opalina
(protozoa)



5 μm

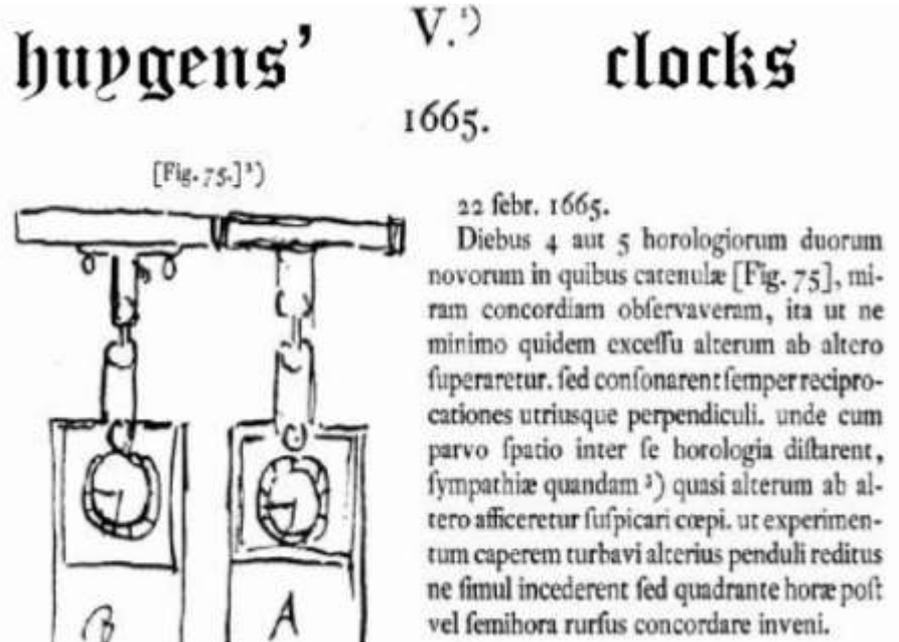
Frog respiratory
mucosa



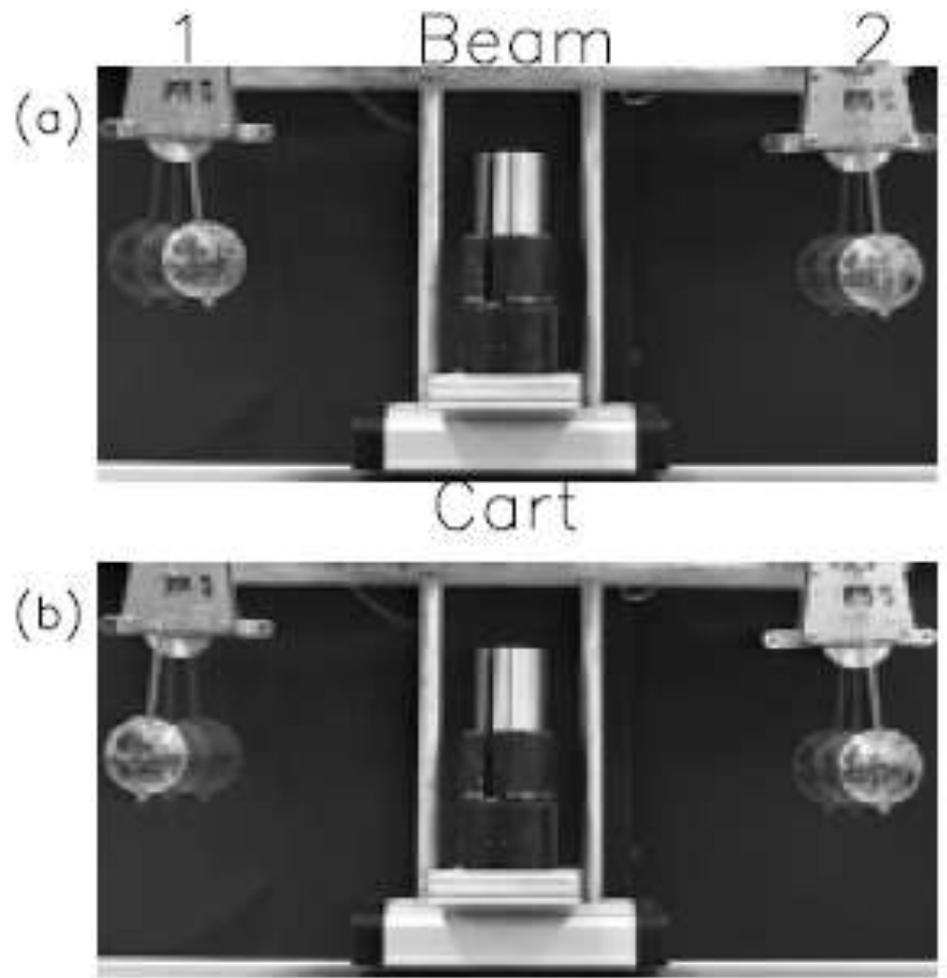
10 μm

Brennen and Winet (1977)

Huygens' Clock Synchronization (1665)



Pendulum clocks hung on a common wall synchronize out of phase!



Modern version of experiment confirms that vibrations in the wall cause the synchronization.

Schatz, et al. (Georgia Tech)

Coupled Metronomes (Lancaster University)



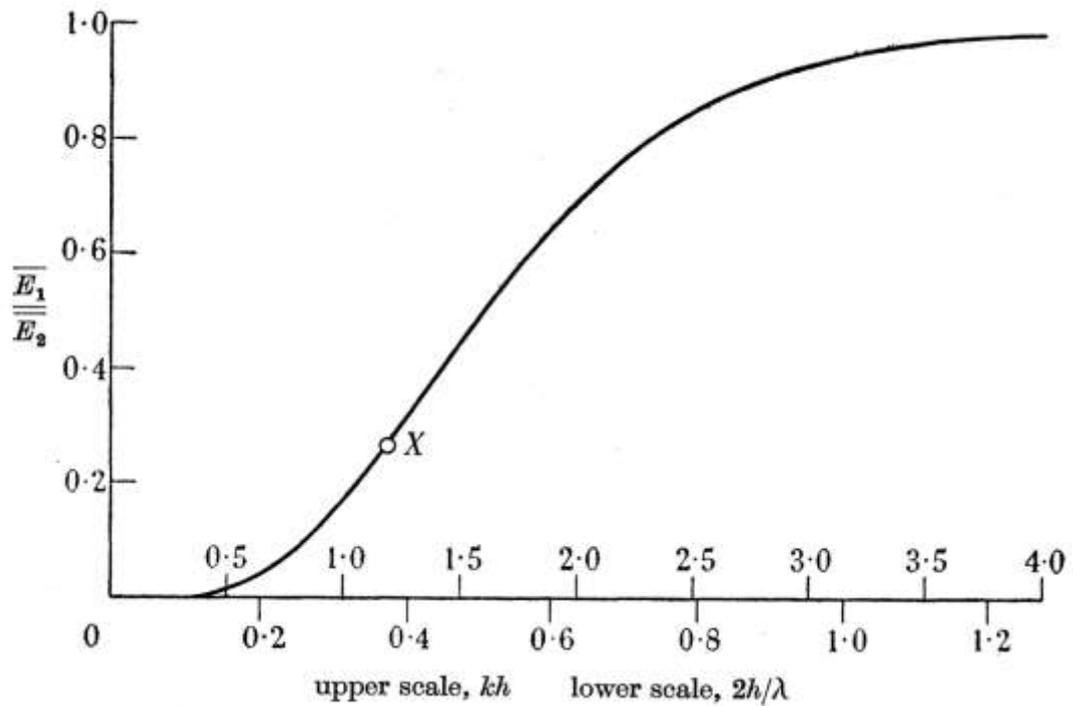
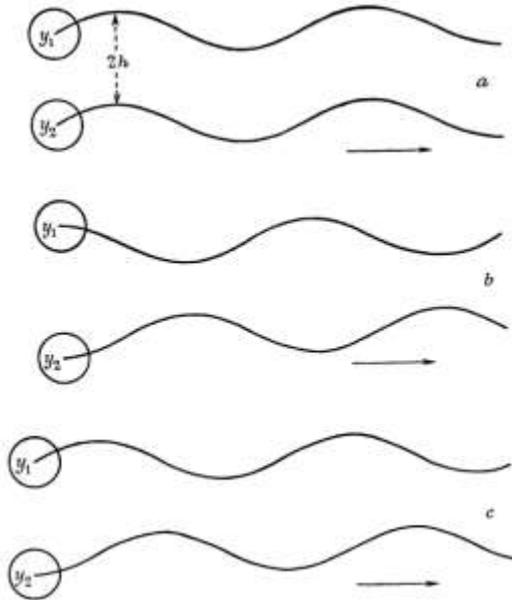
G. I. Taylor's Waving Sheets

Analysis of the swimming of microscopic organisms

BY SIR GEOFFREY TAYLOR, F.R.S.

(Received 25 June 1951)

$\frac{\text{energy dissipation rate in phase}}{\text{energy dissipation rate out of phase}}$



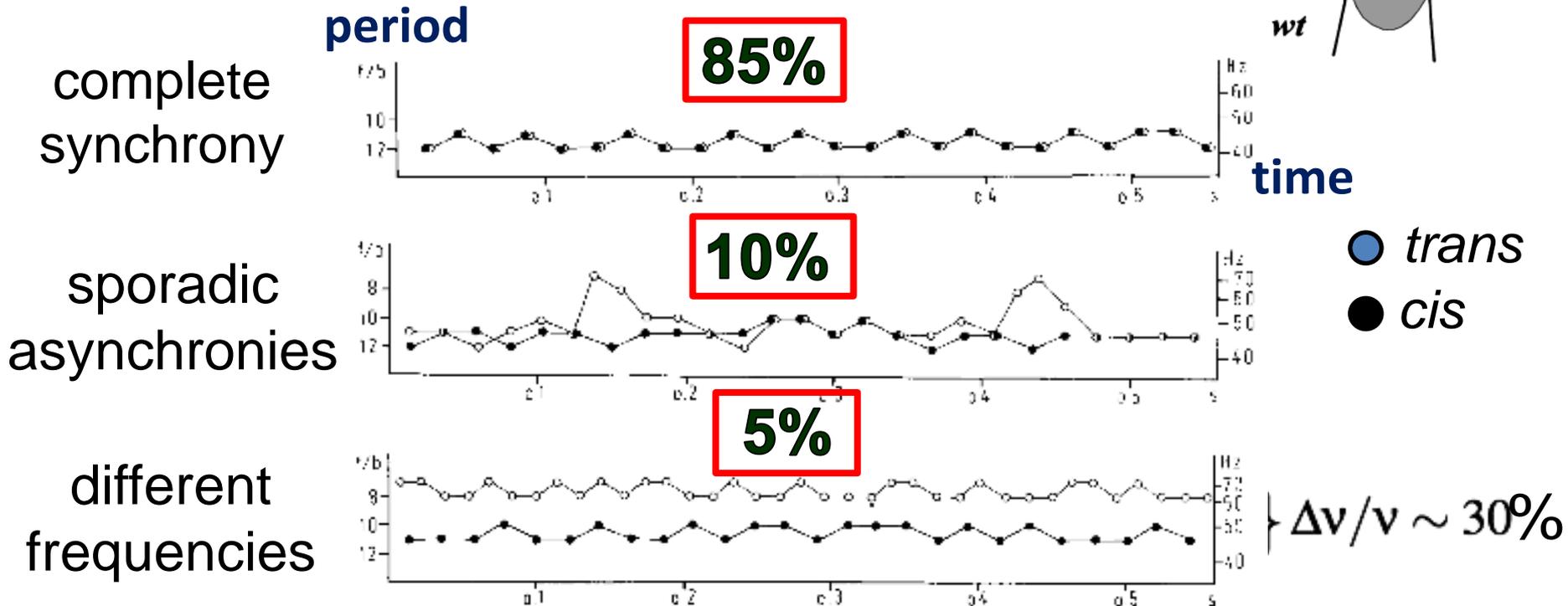
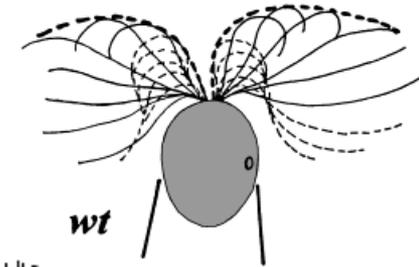
Sheets at various phase relative phase shifts

Scaled separation between sheets

Minimization of dissipation is not (in general) a principle from which to deduce dynamical behaviour...

Early Study of Flagella Synchronisation in *Chlamydomonas*

For different cells:



Attributed to distinct sub-populations of cells

Rüffer and Nultsch, *Cell Motility and the Cytoskeleton* **7**, 87 (1987)

See also: Josef, Saranak, Foster, *Cell Motility and the Cytoskeleton* **61**, 83 (2005)

Historical Background

- R. Kamiya and E. Hasegawa [*Exp. Cell. Res.* ('87)]
(cell models – demembranated)
intrinsically different frequencies of two flagella
- U. Rüffer and W. Nultsch [*Cell Motil.* ('87,'90,'91,'98)]
short observations (50-100 beats at a time, 1-2 sec.)
truly heroic – hand drawing from videos
synchronization, small phase shift, occasional “slips”

Key issue:
control of
phototaxis

“Phase oscillator” model used in e.g. circadian rhythms, etc.

strokes of
flagella

$$S_1(t) = A_1 \cos[\theta_1(t)]$$
$$S_2(t) = A_2 \cos[\theta_2(t)]$$

amplitudes

“phases”
or angles

$$\frac{d\theta_1}{dt} = \omega_1 + \dots$$
$$\frac{d\theta_2}{dt} = \omega_2 + \dots$$

natural
frequencies

Without coupling, the phase difference simply grows in time

$$\Delta \equiv \frac{\theta_1 - \theta_2}{2\pi} = (\nu_1 - \nu_2)t + \dots$$

So, is this seen?

Noisy Synchronization

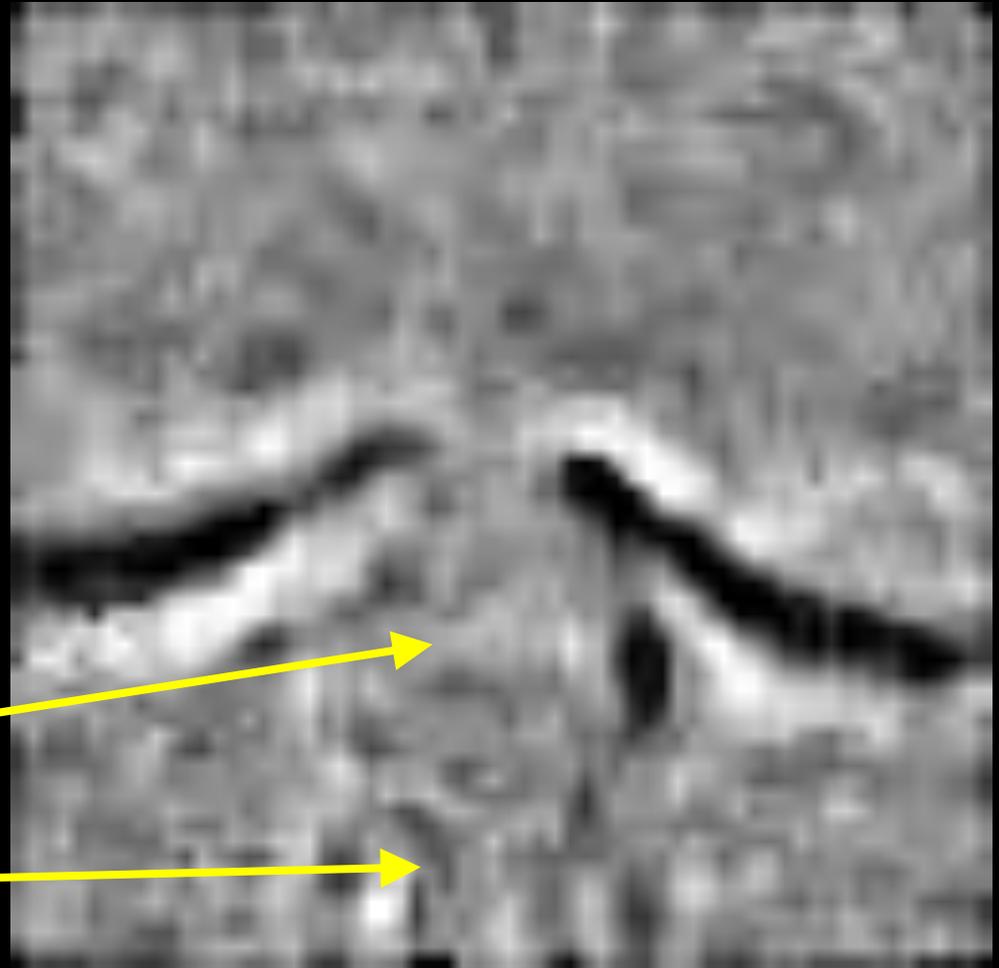
Frame-subtraction

Experimental methods:

- Micropipette manipulation with a rotating stage for precise alignment
- Up to 2000 frames/sec
- Long time series (50,000 beats or more)
- Can impose external fluid flow

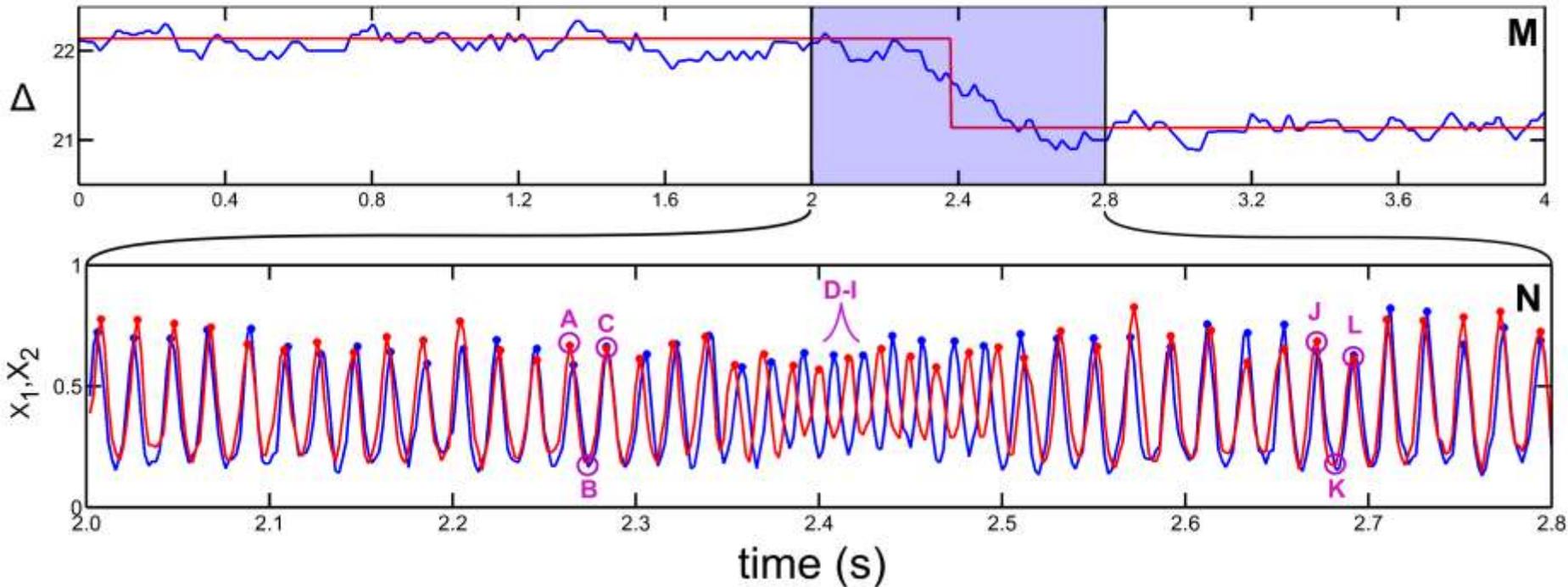
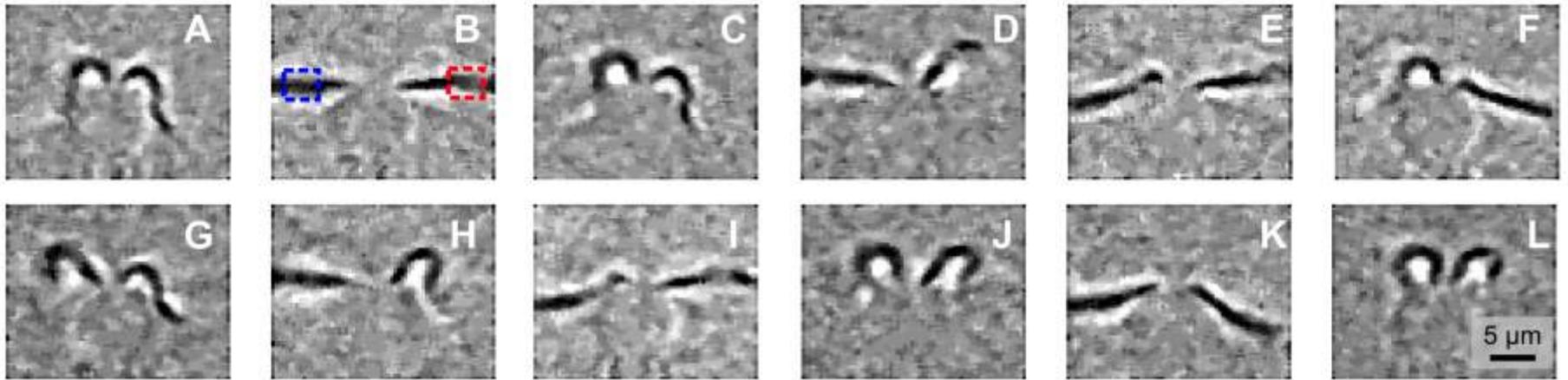
Cell body

Micropipette



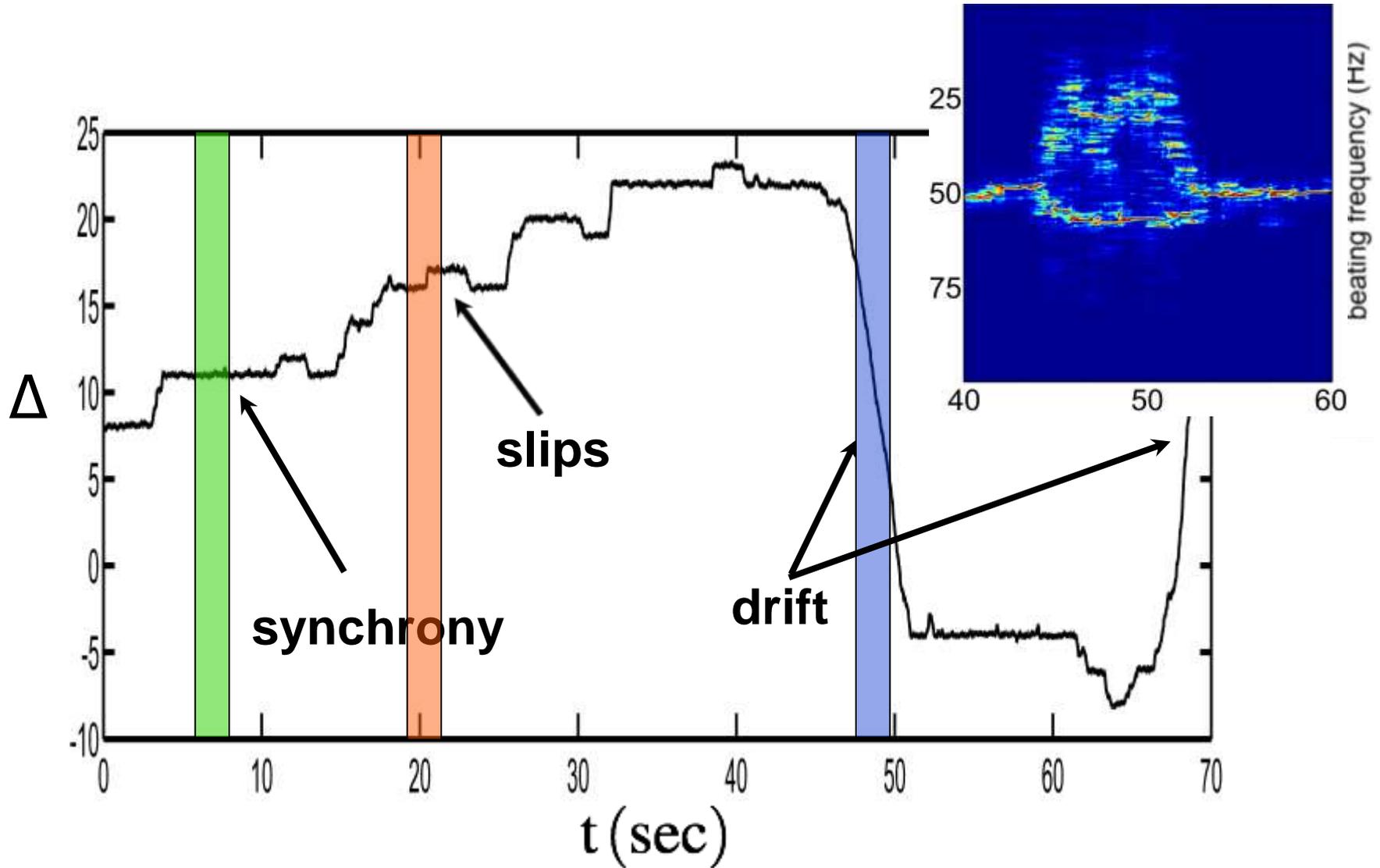
Polin, Tuval, Drescher, Gollub, Goldstein, *Science* **325**, 487 (2009)

A Phase Slip



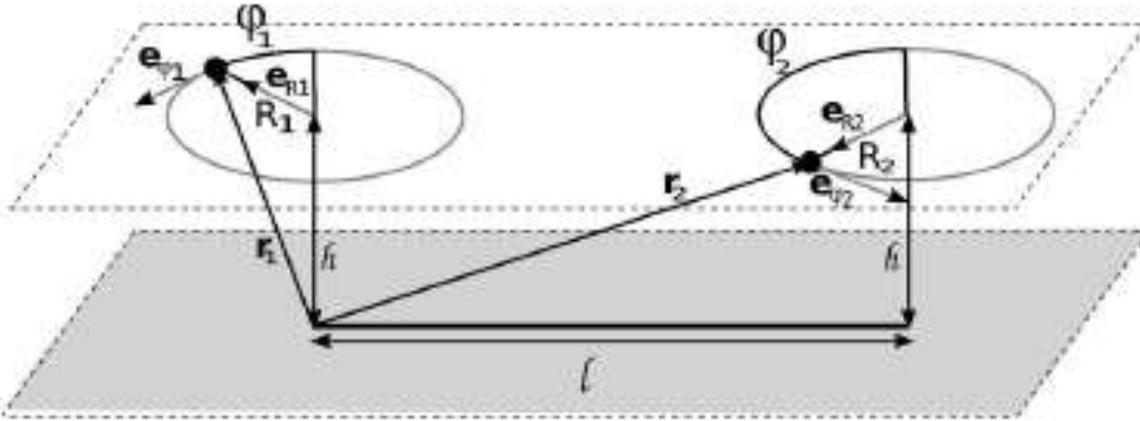
Goldstein, Polin, Tuval, *Phys. Rev. Lett.* **103**, 168103 (2009)

A Single Cell Displays All Three Regimes (!)



Polin, Tuval, Drescher, Gollub, Goldstein, *Science* **325**, 487 (2009)

Model for Phase Evolution



Spheres forced in circular orbits by an azimuthal force, with elasticity to maintain orbit radius, and sphere-sphere hydrodynamic interactions (*deterministic*)

Niedermayer, Eckhardt, and Lenz, *Chaos* (2008)
 See also: Guirao and Joanny, *Biophys. J.* (2007)
 Vilfan and Julicher, *PRL* (2006)



We see clear evidence of stochasticity ...
 which suggests the stochastic **Adler** equation:

$$\dot{\Delta} = \delta\nu - 2\pi\varepsilon \sin(2\pi\Delta) + \xi(t)$$

Intrinsic frequency mismatch (hydrodynamics?)

coupling Strength

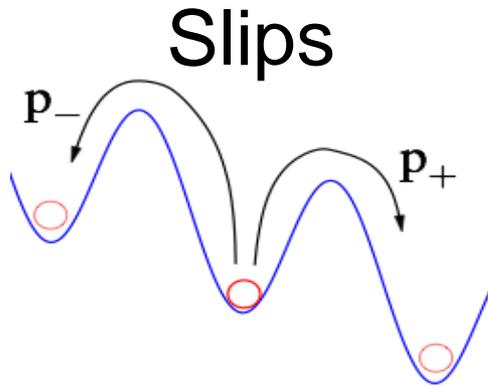
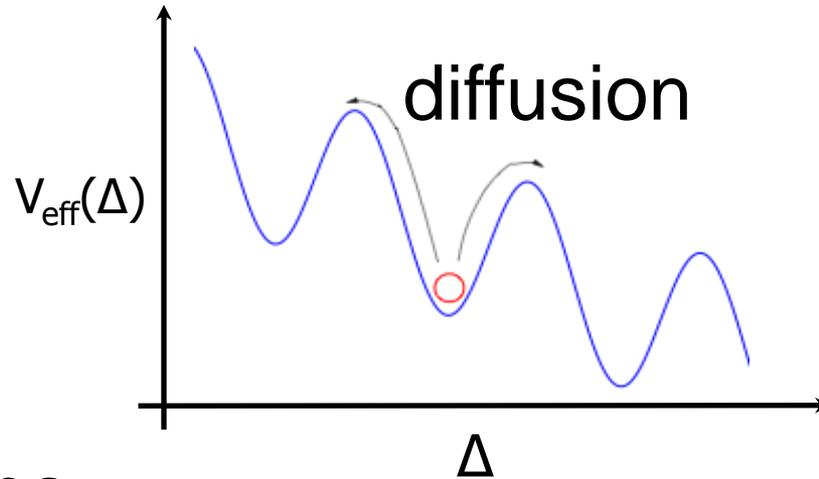
Quasi-universal form for phase oscillators (Kuramoto)

biochemical noise

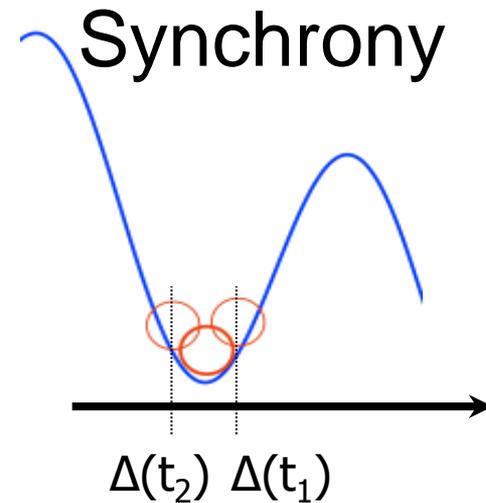
$$\langle \xi(t) \rangle = 0$$

$$\langle \xi(t)\xi(s) \rangle = 2T_{eff} \delta(t-s)$$

Model for Phase Evolution

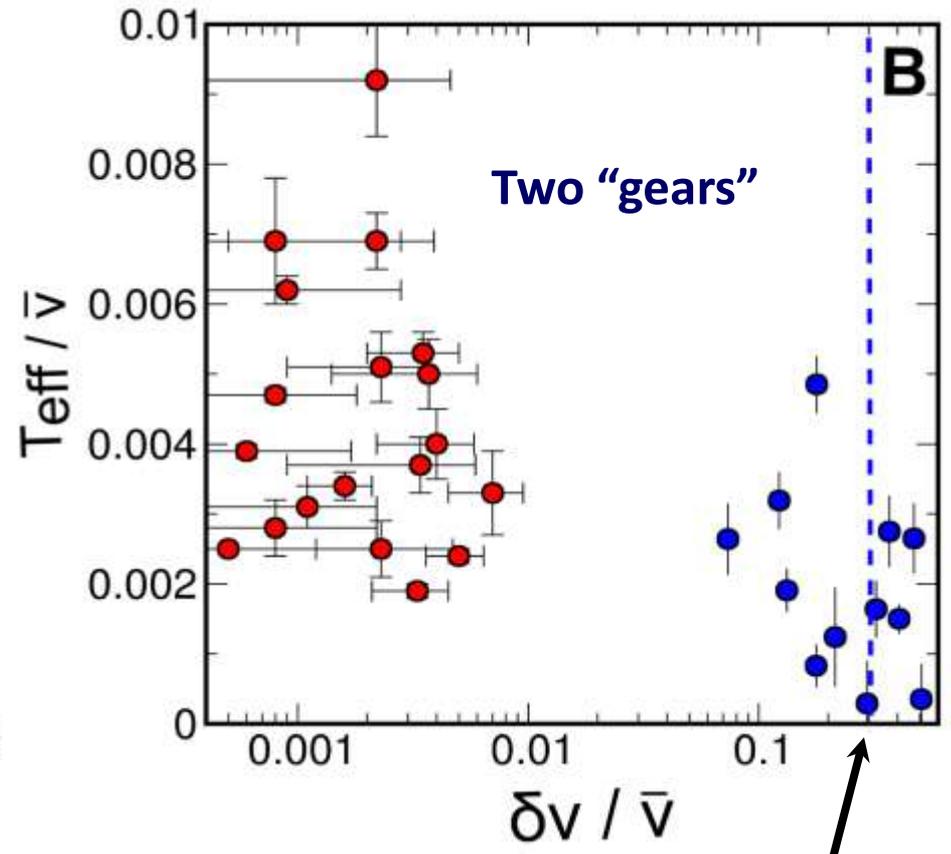
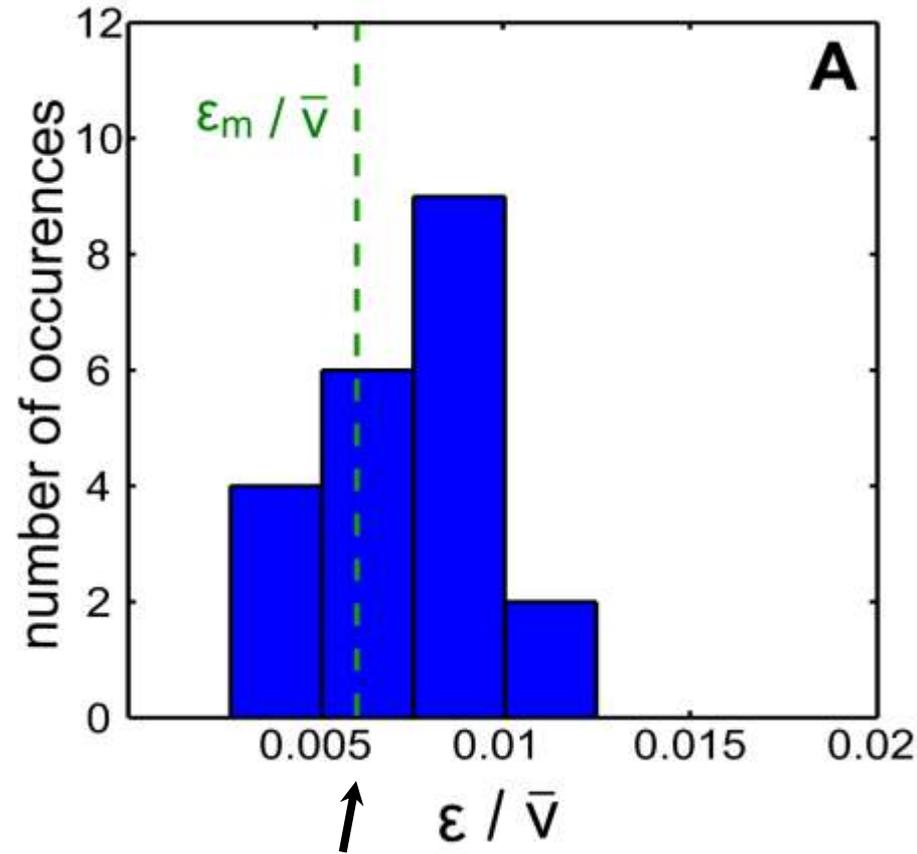


Relative probability of +/- slips
Yields the frequency difference $\delta\nu$



Amplitude and autocorrelation
function of fluctuations in the
synchronised state yields T_{eff} and ε

Model Parameters



estimate of hydrodynamic coupling

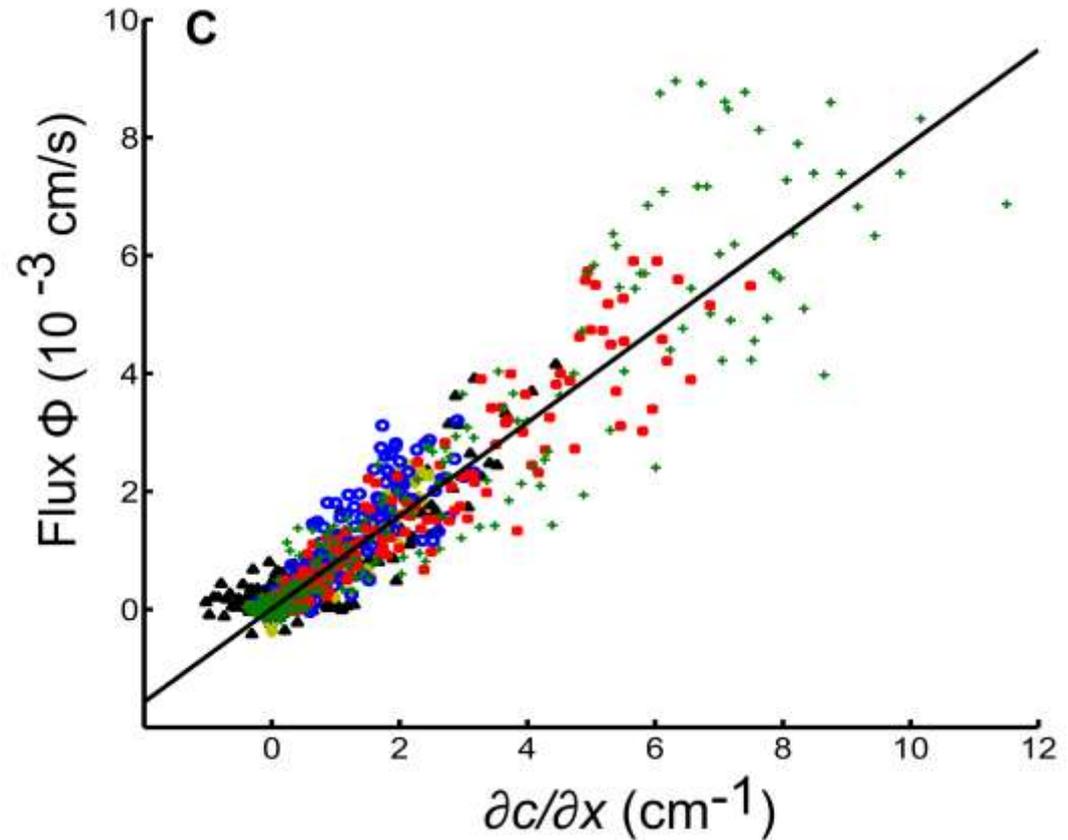
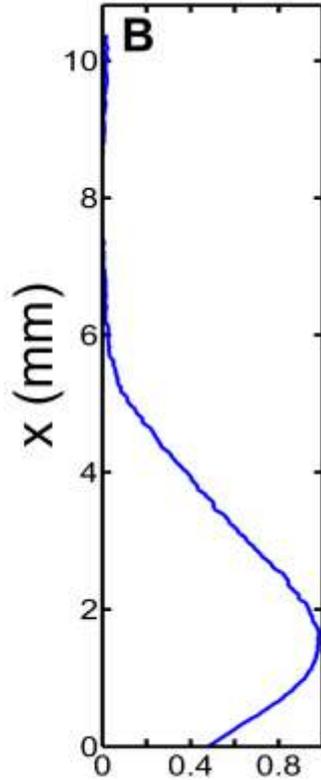
$$\epsilon \approx \tau_r V_L V_R$$

$$\tau_r = \frac{3\pi\eta a l^3}{\kappa}$$

expected value for intrinsic frequency difference

Polin, *et al.*, *Science* **325**, 487 (2009)

Direct Demonstration of *Chlamydomonas* Diffusion

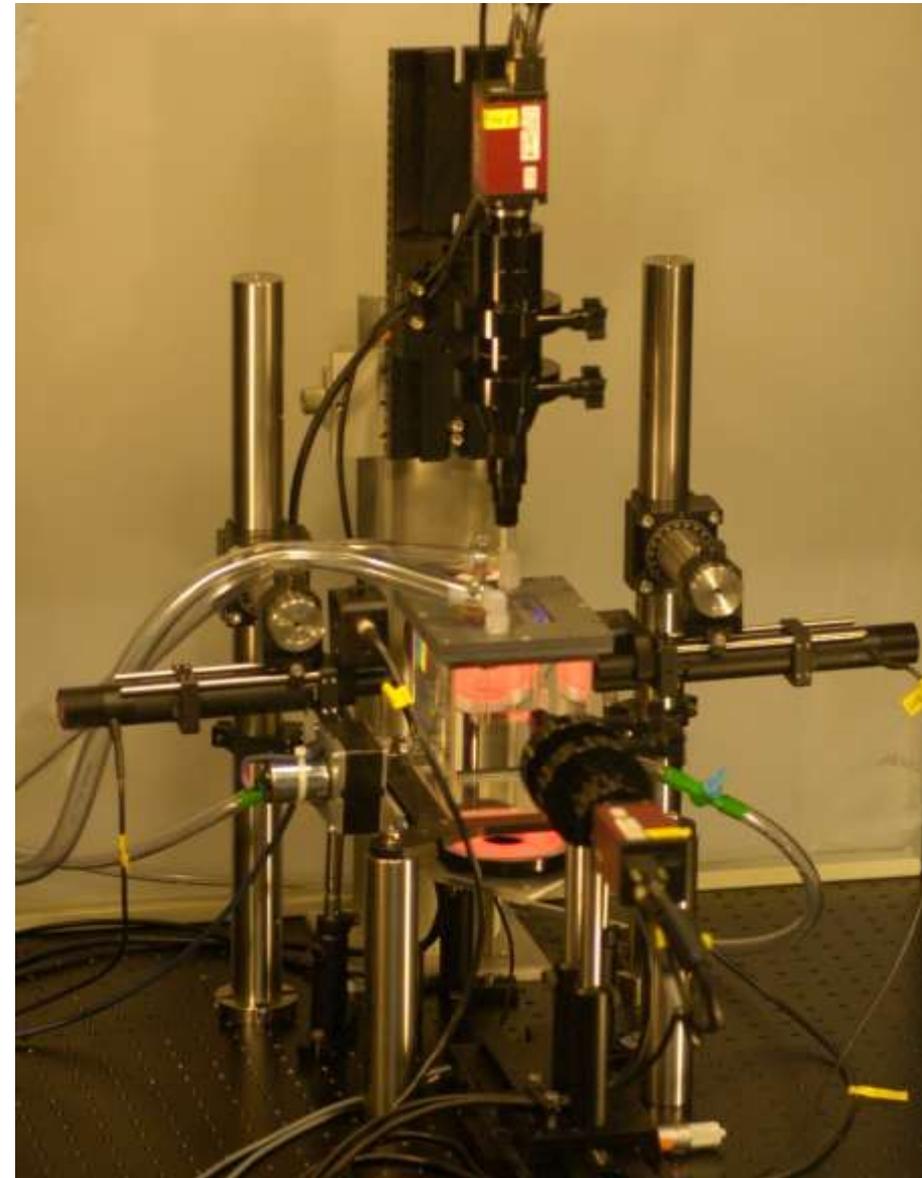
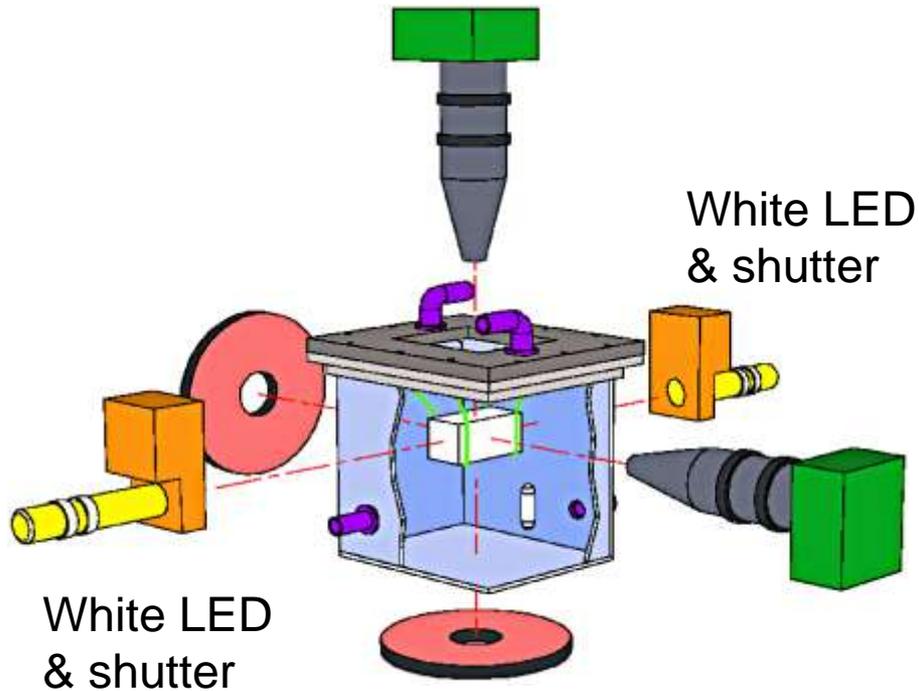


$$D_{\text{exp}} \sim (0.68 \pm 0.11) \times 10^{-3} \text{ cm}^2/\text{s}$$

Since $D \sim u^2 \tau$ and $u \sim 100 \mu\text{m}/\text{s}$, there must be a time $\tau \sim 10 \text{ s}$

Polin, Tuval, Drescher, Gollub, Goldstein, *Science* **325**, 487 (2009)

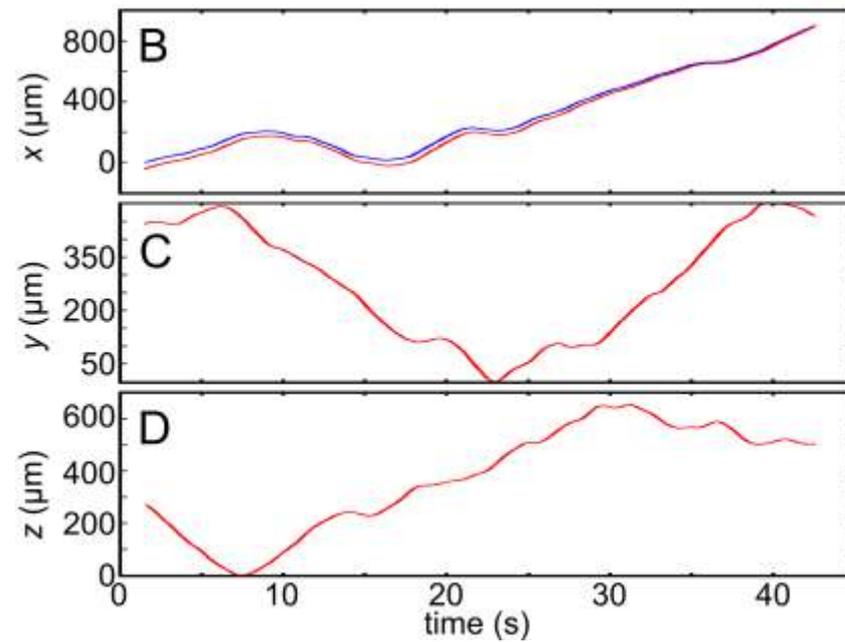
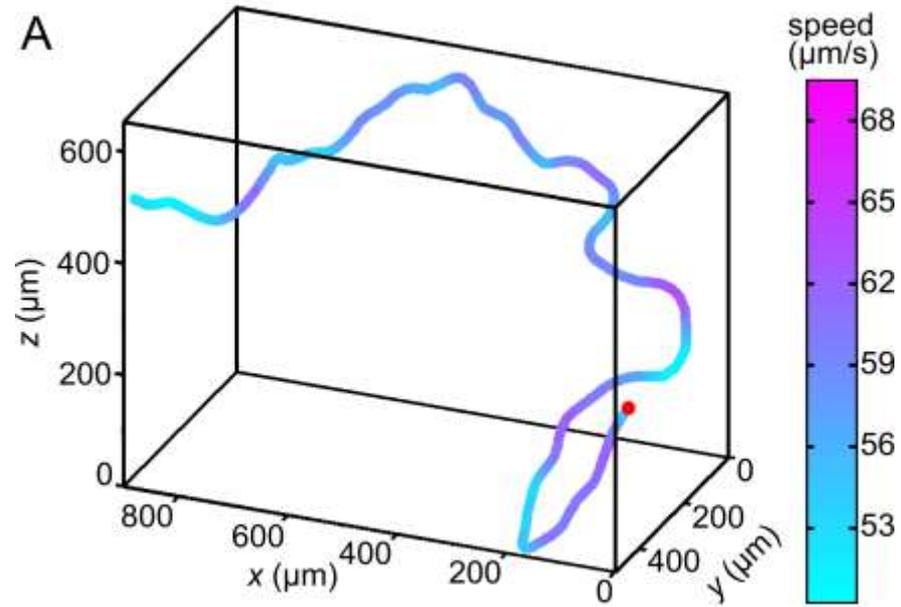
Dual-View Apparatus Free of Thermal Convection



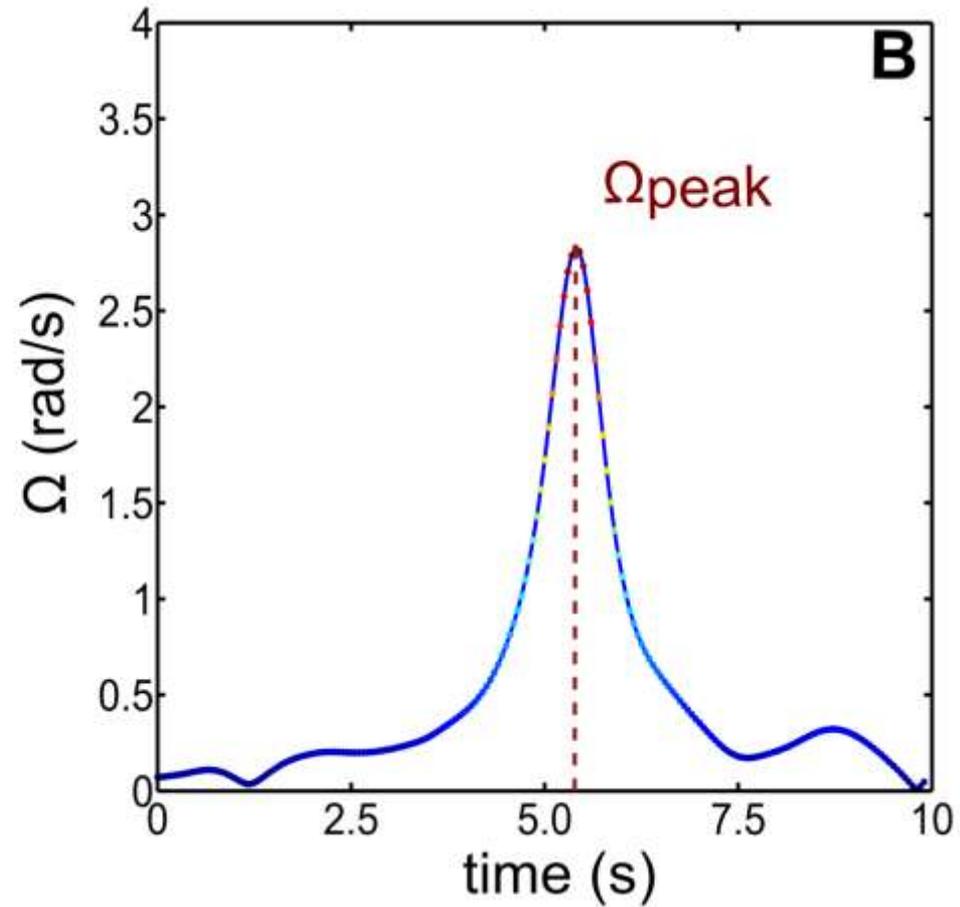
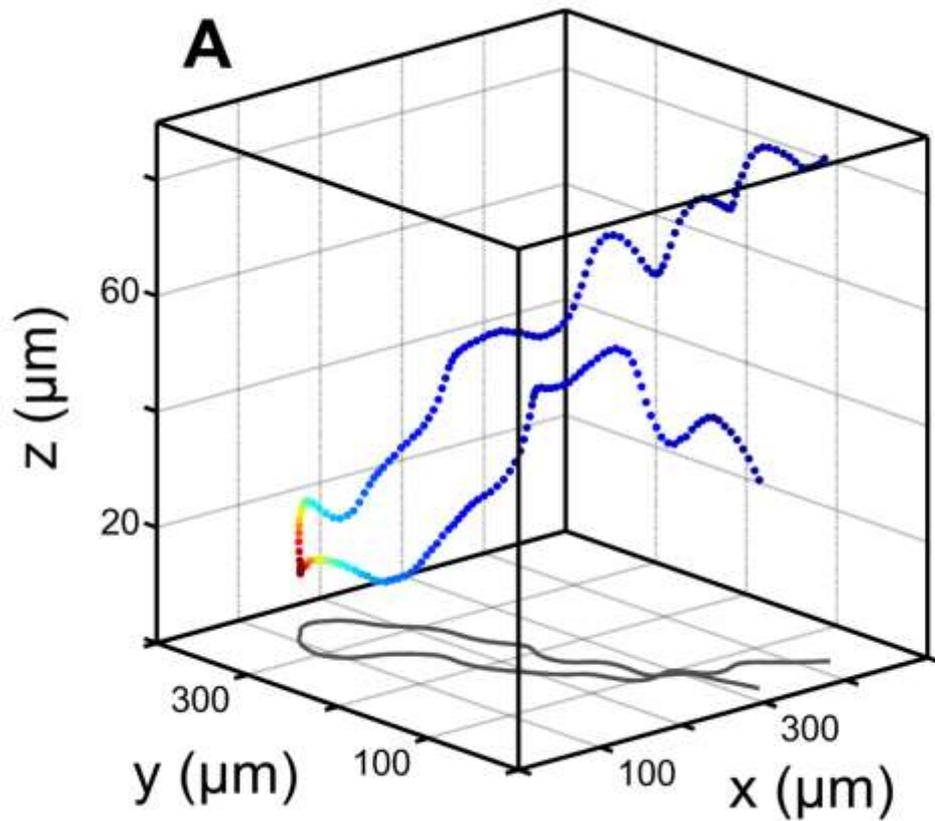
Capable of imaging protists from $10\ \mu\text{m}$ to 1 mm, with tracking precision of ~ 1 micron, @ 20 fps.

Drescher, Leptos, Goldstein,
Review of Scientific Instruments **80**, 014301 (2009)

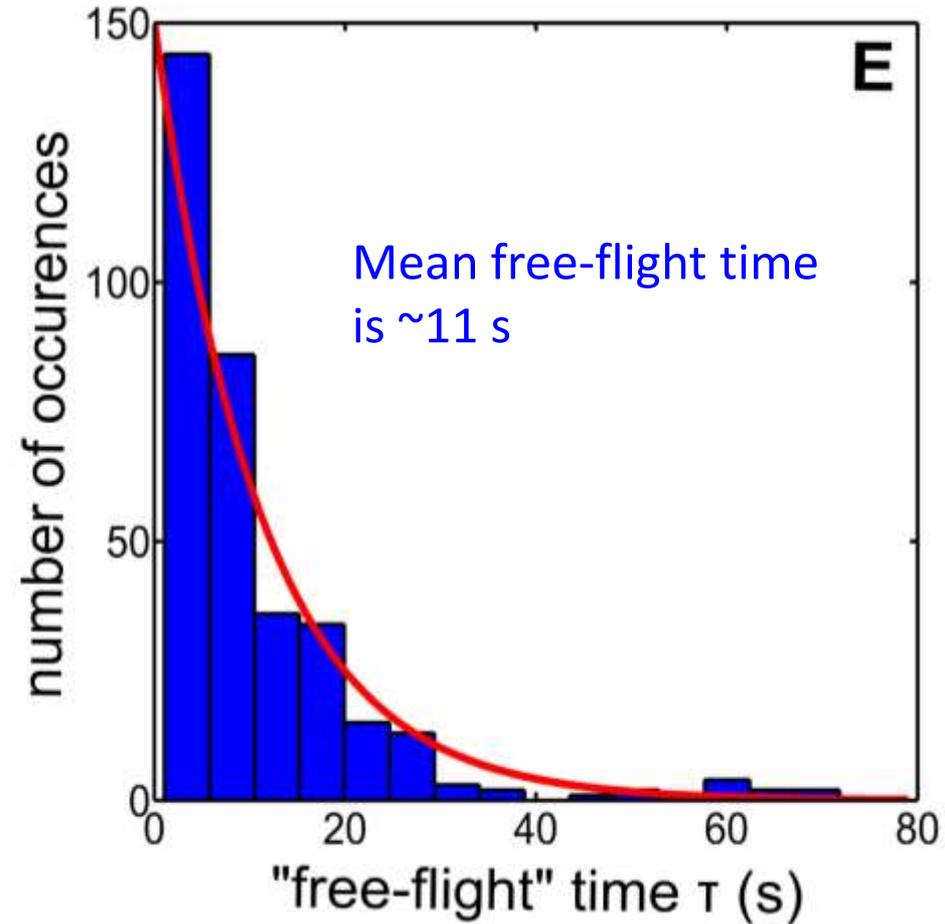
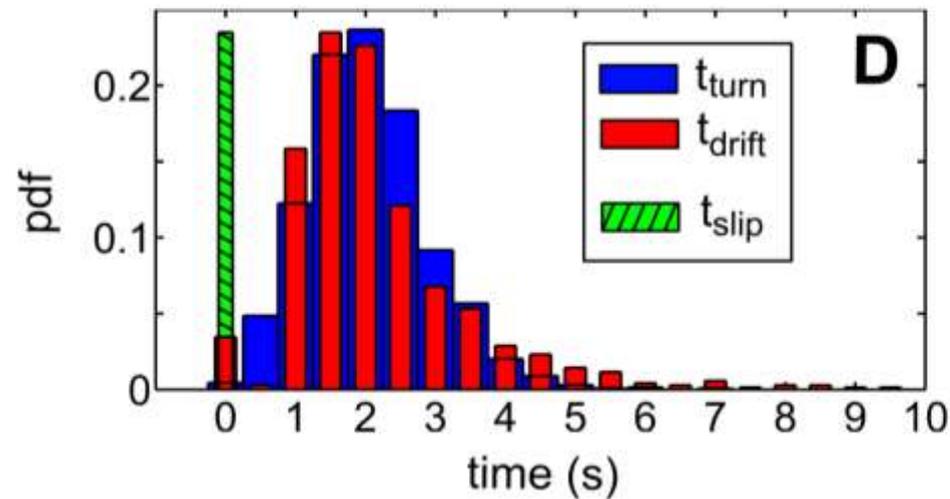
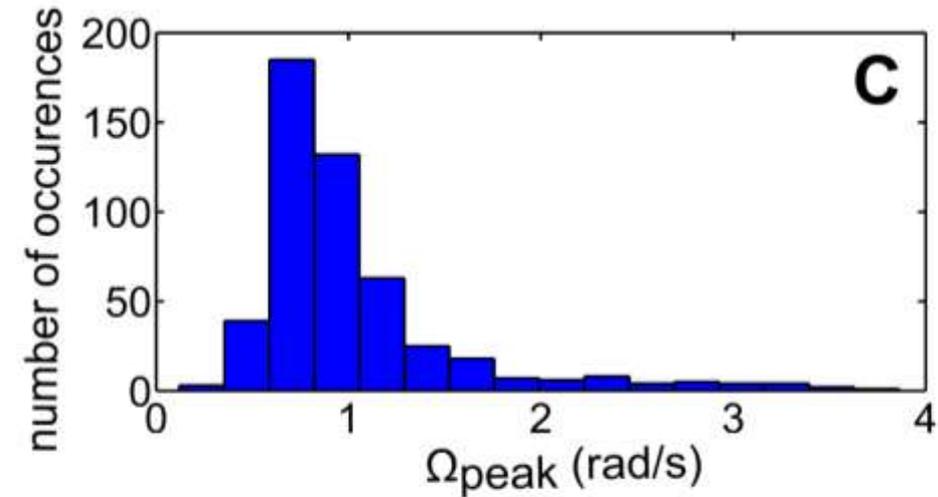
Reconstructing a Trajectory



Chlamydomonas Tracking in Detail – A Sharp Turn



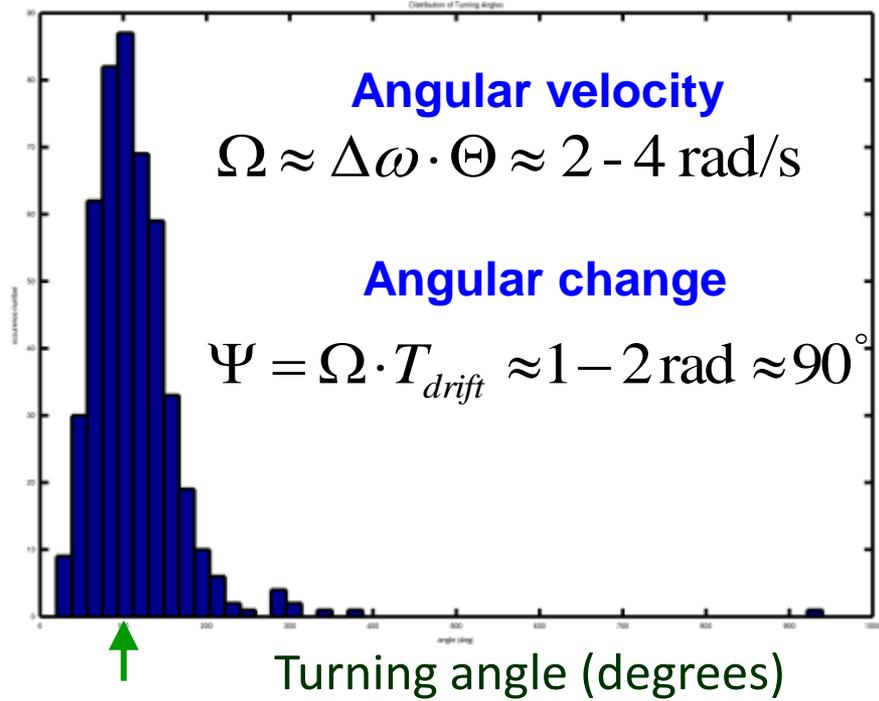
Statistics of Sharp Turns: Origin of Diffusion



Turns and drifts have identical statistics, much longer than slips.

Geometry of Turning

Chlamy w/single flagellum, rotating near a surface



Angle per beat - $\Theta = \frac{2\pi}{\sim 16 \text{ beats}} \approx 0.4 \frac{\text{rad}}{\text{beat}}$

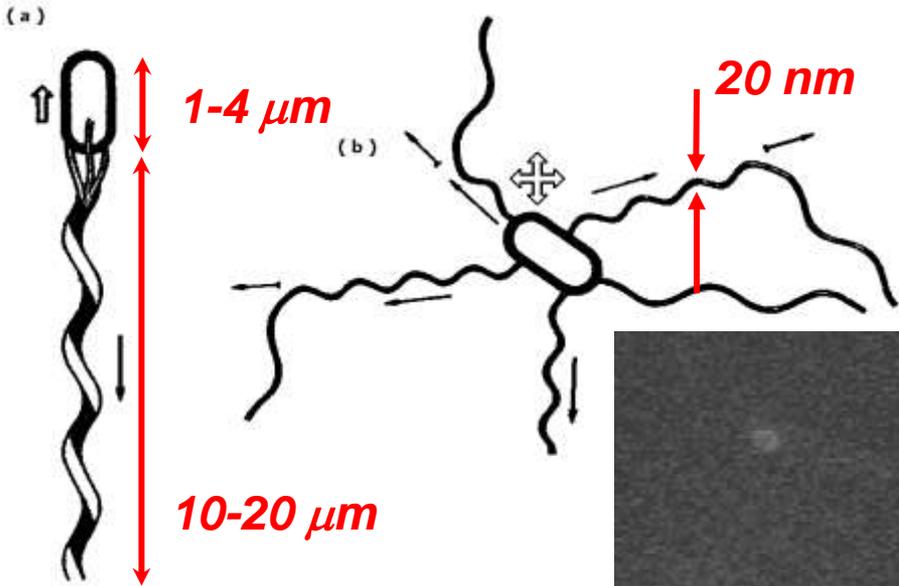
Frequency difference - $\Delta\omega \approx 5 - 10 \frac{\text{beats}}{\text{s}}$

“Drift” duration- $T_{drift} \approx 25 - 50 \text{ beats}$
 $\approx 0.5 - 1 \text{ s}$

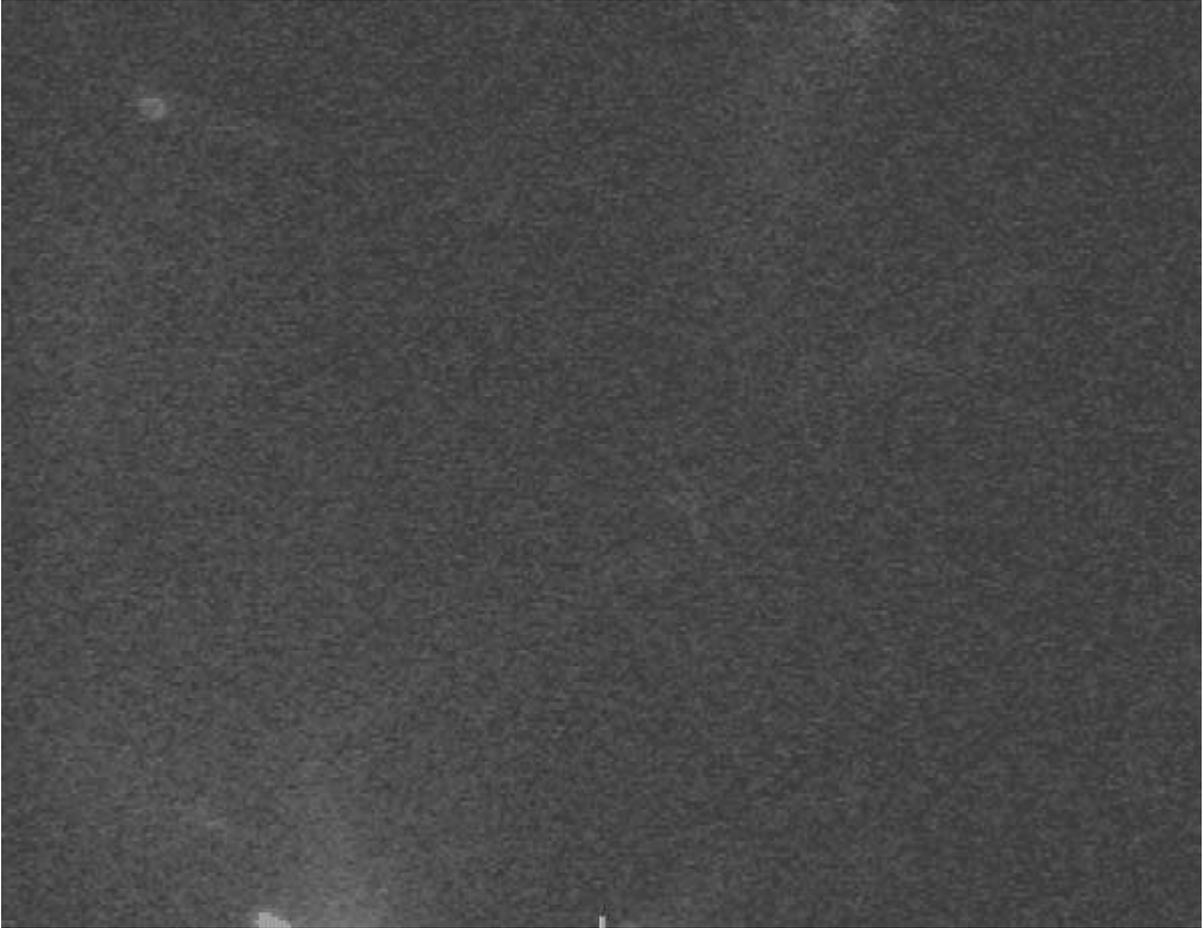
$$D = \frac{u^2 \tau}{3(1 - \cos\theta)}$$

$D_{est} \sim (0.47 \pm 0.05) \times 10^{-3} \text{ cm}^2/\text{s}$

Run-and-Tumble Locomotion of Bacteria

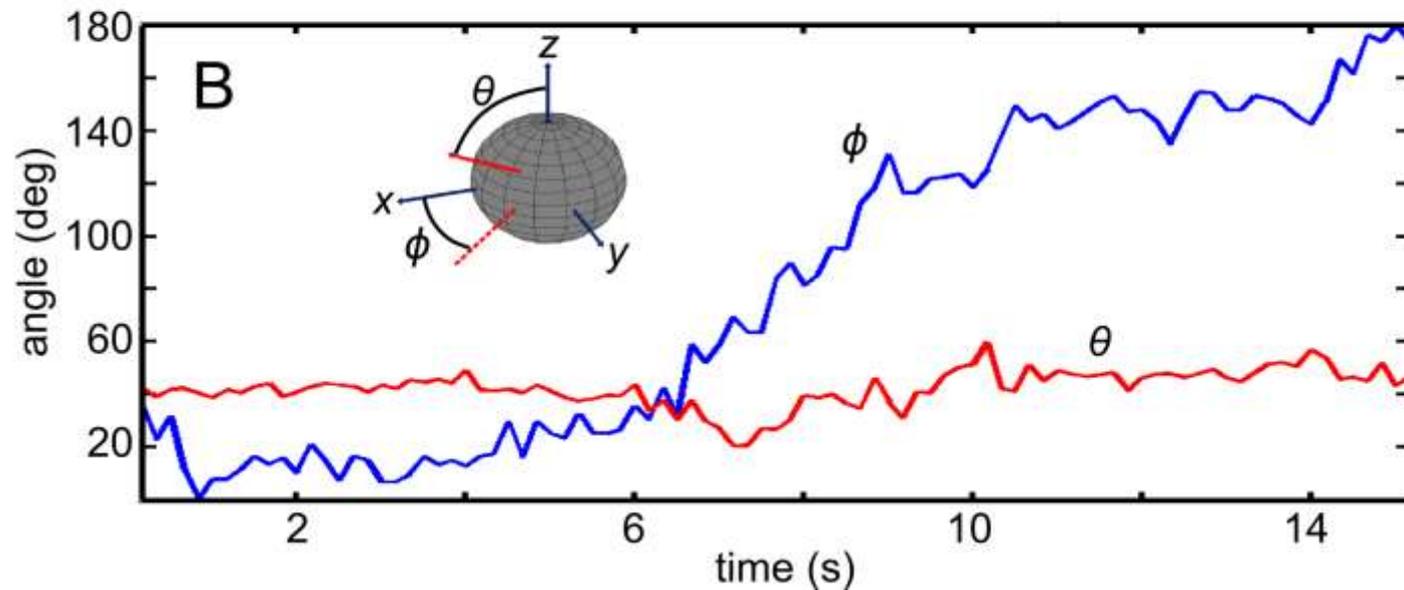
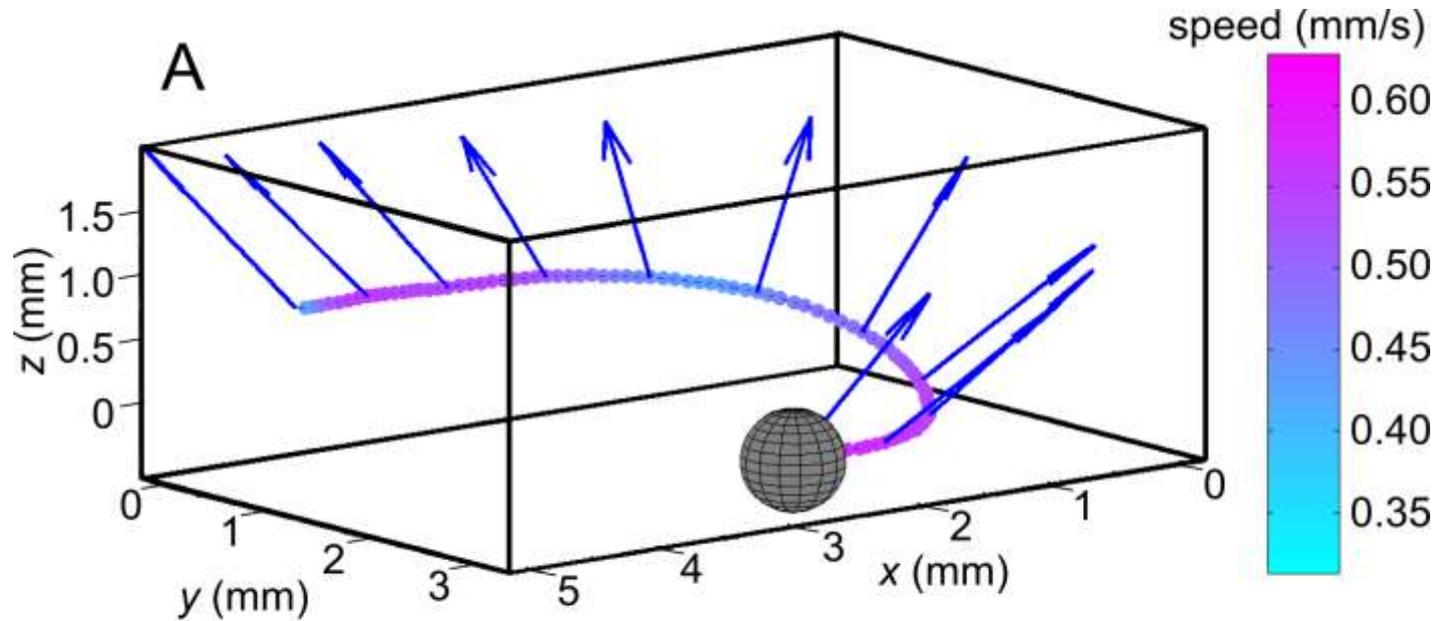


Turner, Ryu, and Berg (2000)



Macnab and Ornstein (1977)

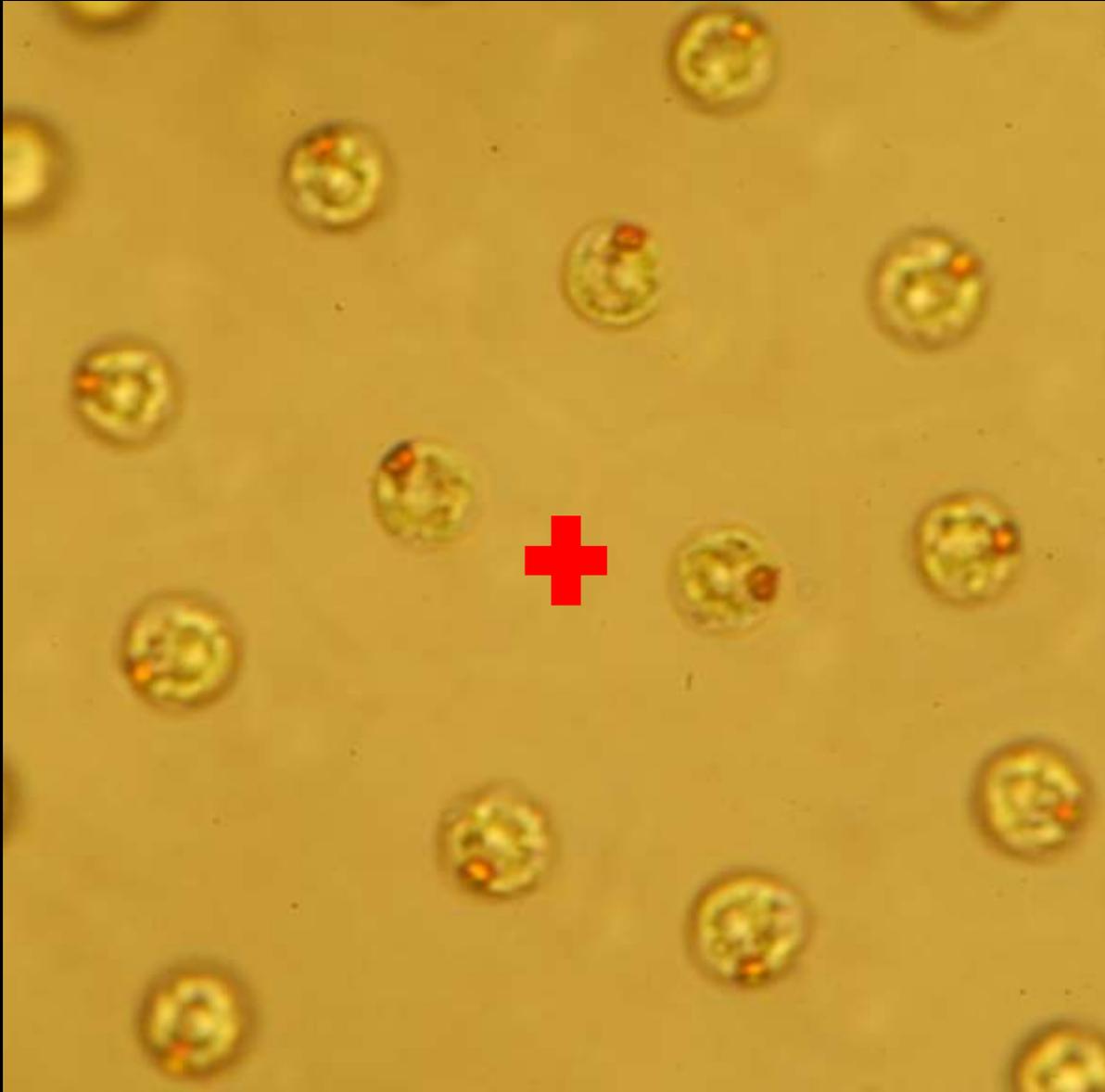
A Phototurn (*V. barberi*) With Bottom-Heaviness



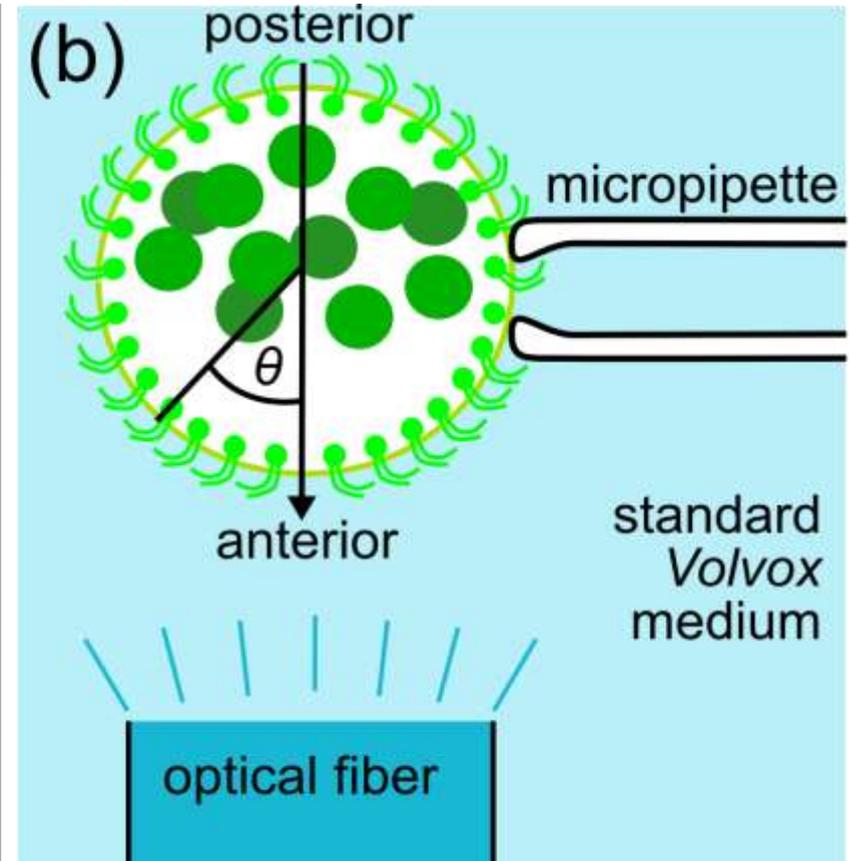
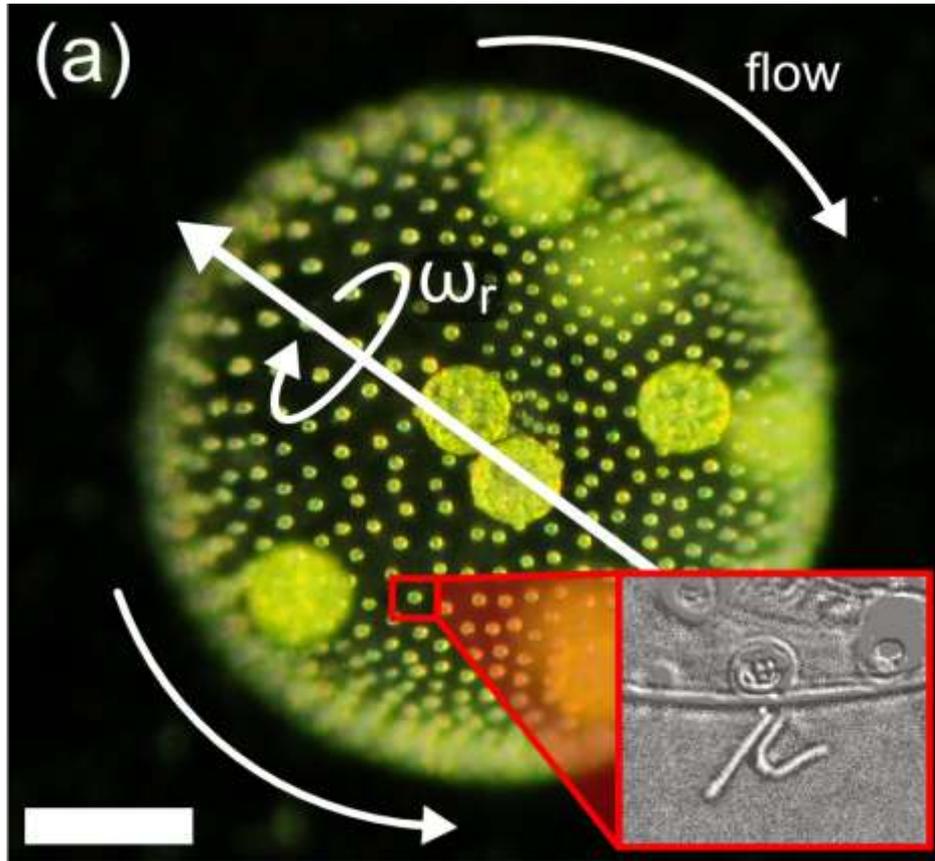
Drescher, Leptos, Goldstein, *Rev. Sci. Instrum.* (2009)

Volvox Eyespots

Top view at anterior pole

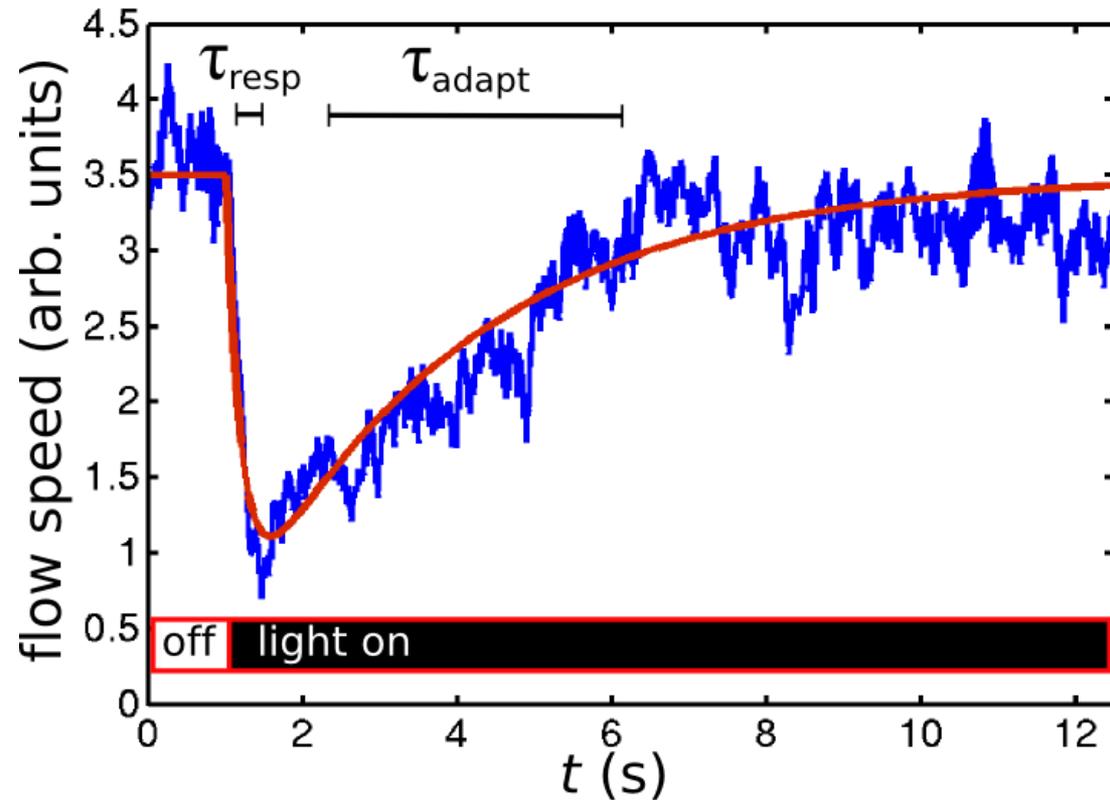


Adaptive Flagellar Dynamics and the Fidelity of Multicellular Phototaxis



Drescher, Goldstein, Tuval, *preprint* (2010)

Dynamic PIV Measurements – Step Response



Adaptive dynamics also play a role in sperm chemotaxis:
Friedrich and Jülicher (2007,09)

Adaptive, two-variable model

$$\dot{p} = \frac{(s - h) - p}{\tau_r}$$

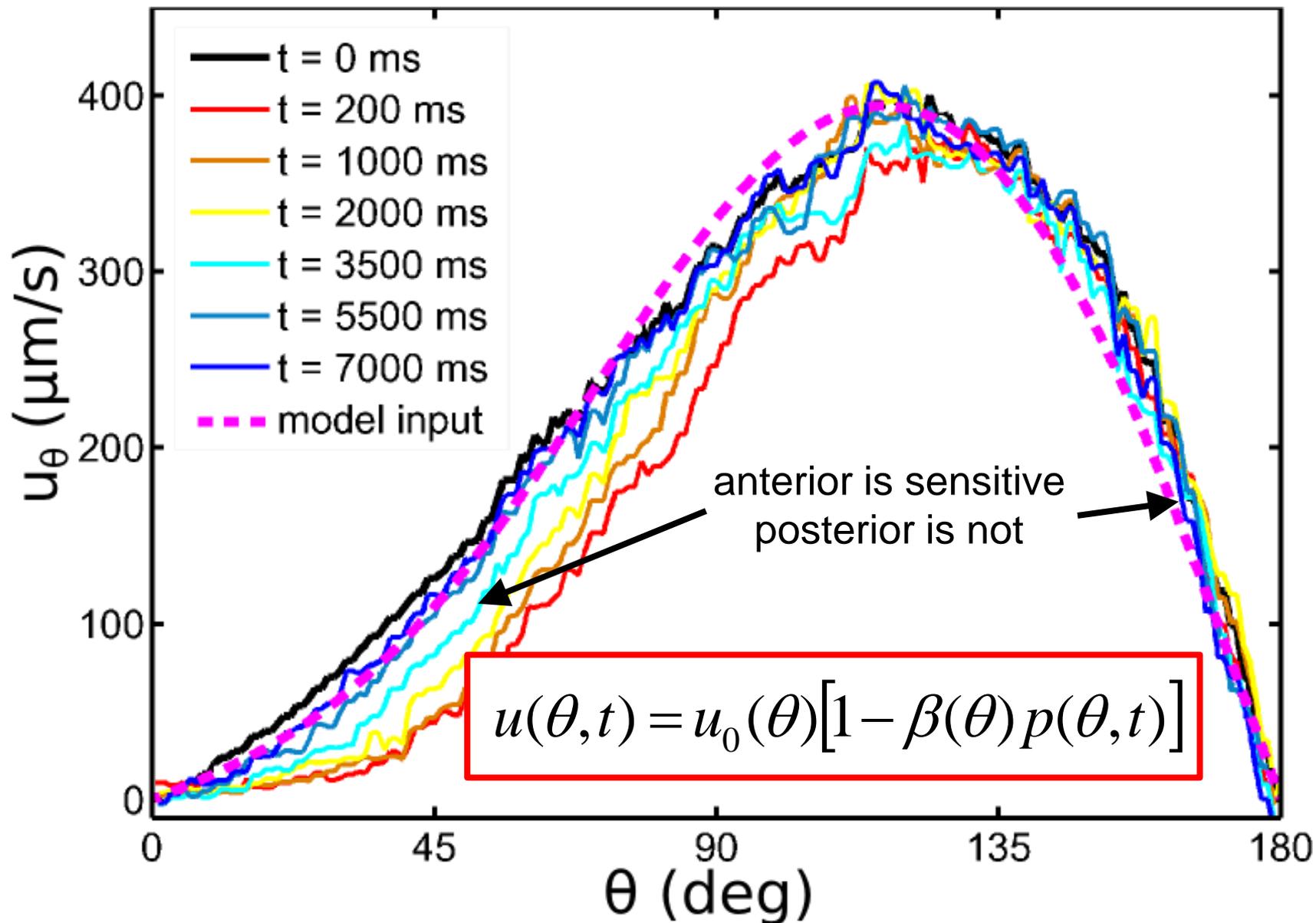
$$\dot{h} = \frac{s - h}{\tau_a}$$

p = "photoresponse" amplitude
 h = "hidden" biochemistry

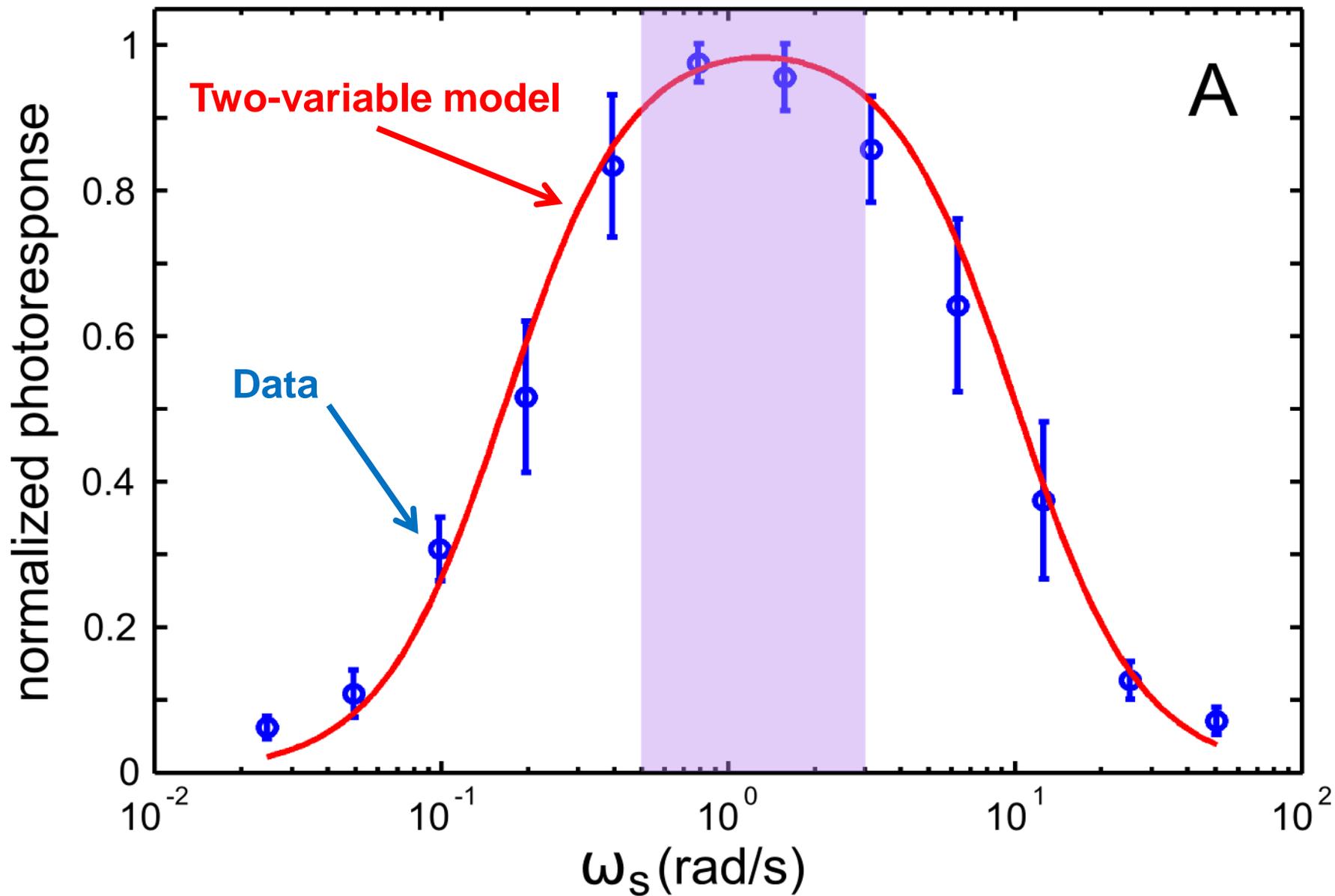
$$u = u_0 (1 - \beta p)$$

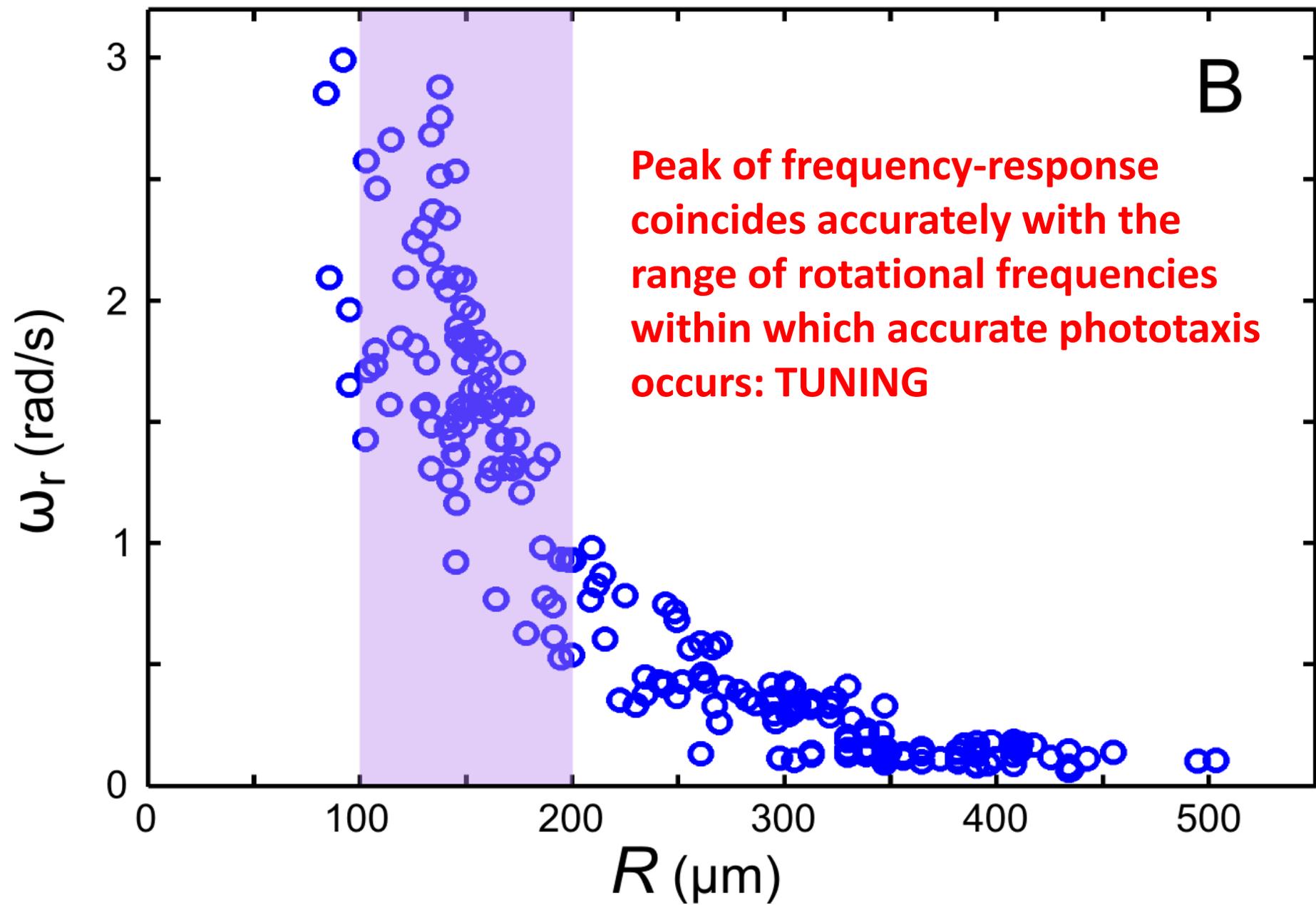
Simple modulation of flow

Angular Dependence of the Transient Response

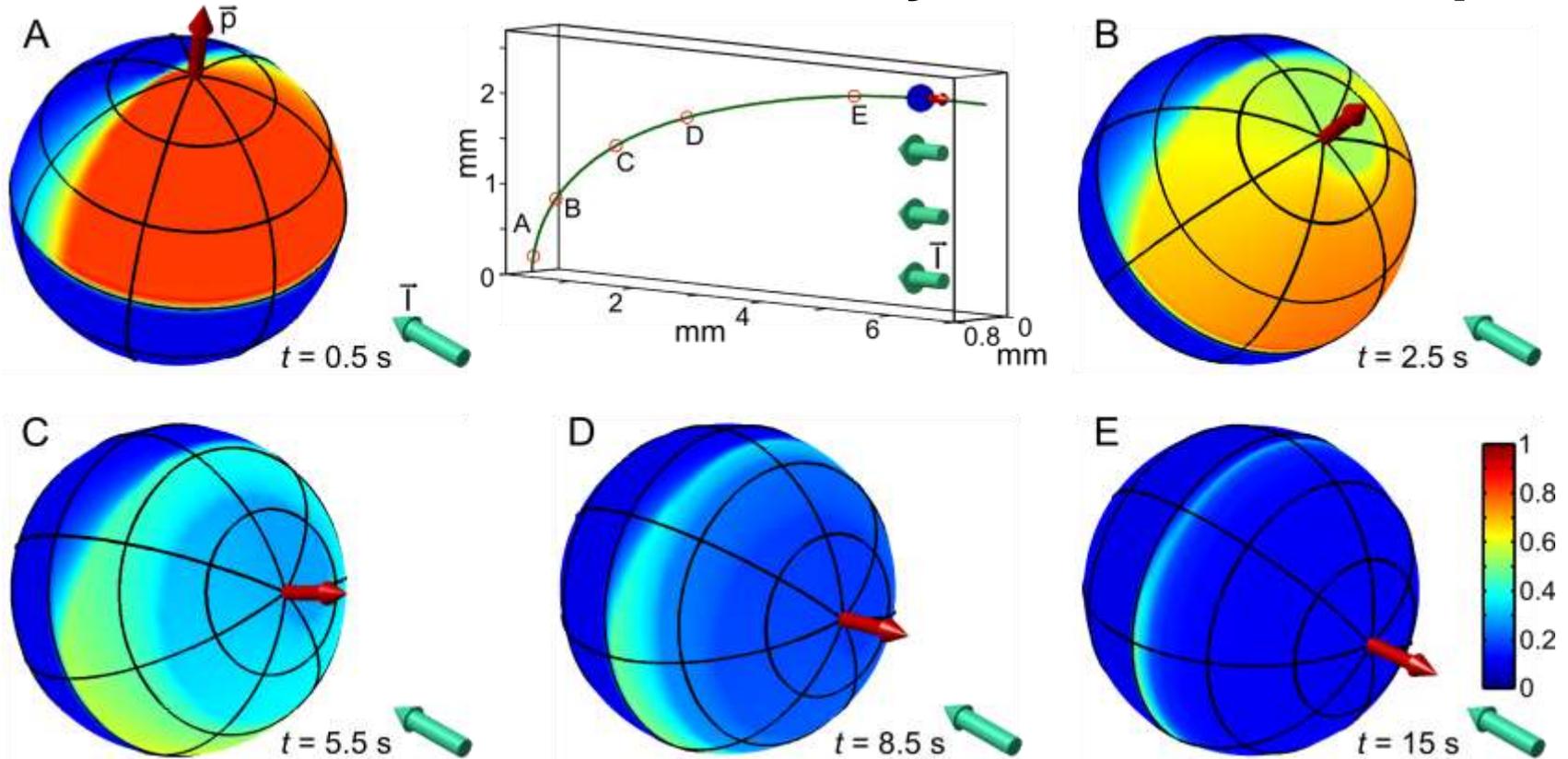


Frequency-Dependent Response





Multicellular Phototaxis as Dynamic Phototropism

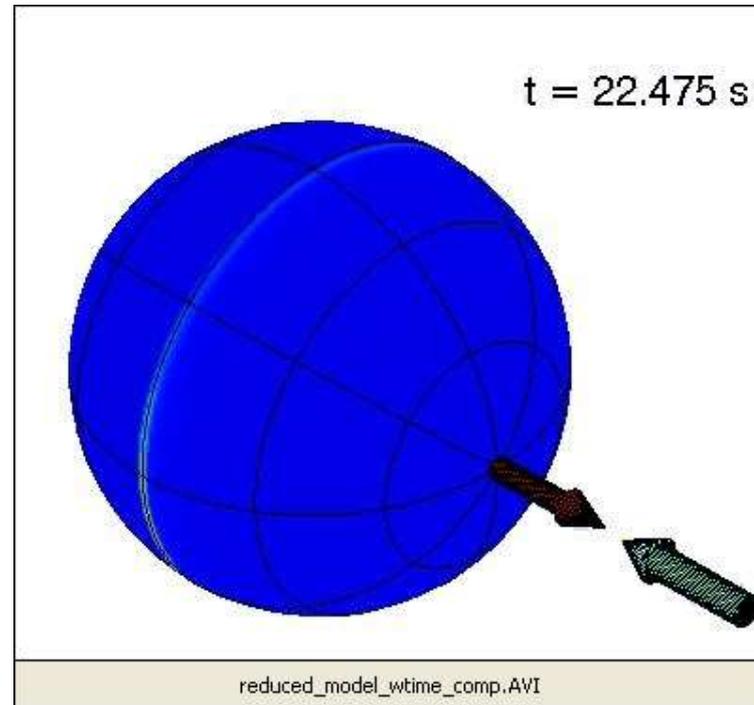


Initial: illuminated anterior region has strongly diminished flagellar beating, colony rotates toward the light

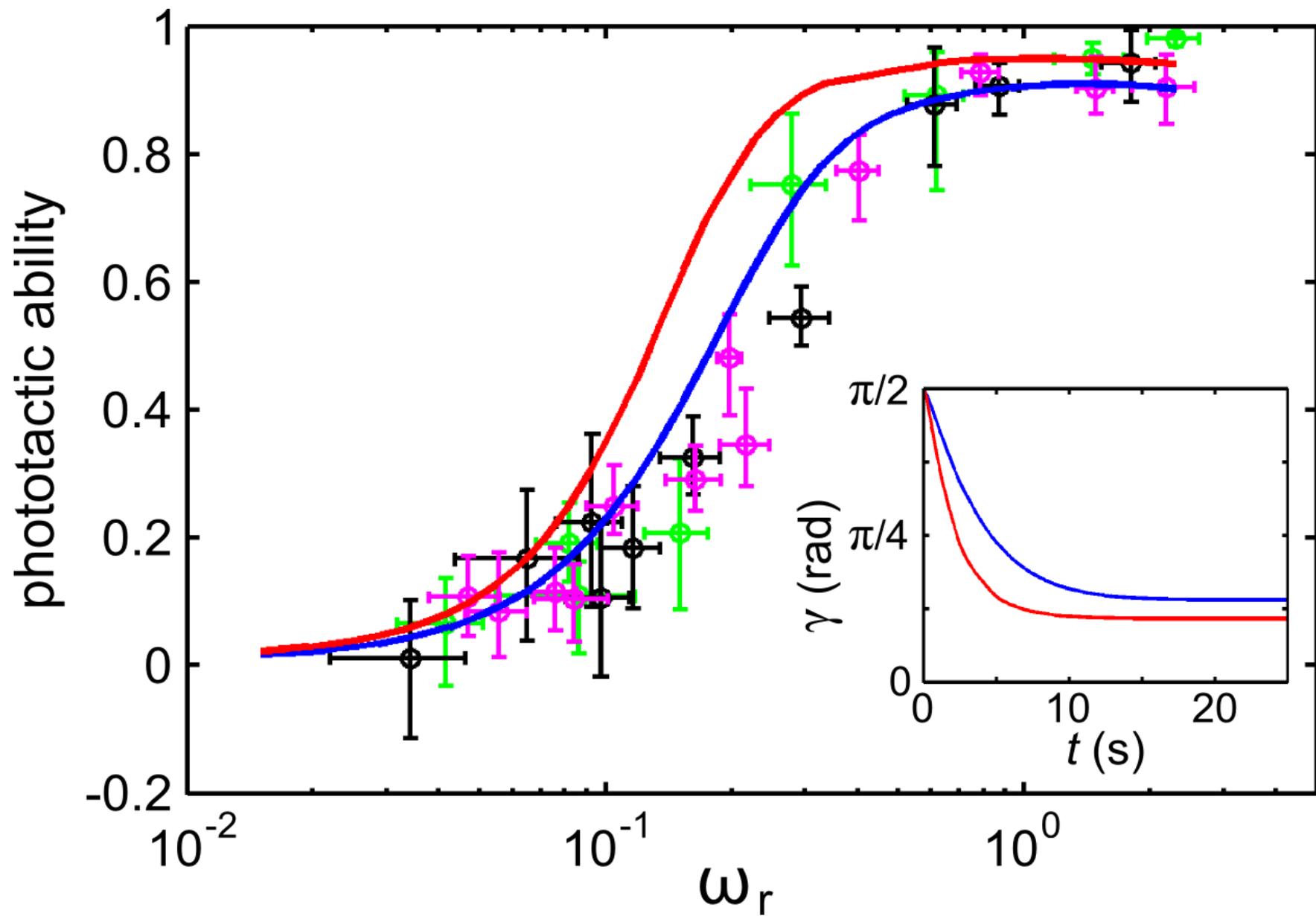
Midway: modulation amplitude decreases as colony axis tilts toward light

Done: no more modulation of flagellar beating, axis aligned

Phototaxis: The Movie



A Test of the Theory



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