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Rotation of slender swimmers in isotropic-drag media

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The drag anisotropy of slender filaments is a critical physical property allowing swimming in low-Reynolds number flows, and without it linear translation is impossible. Here we show that, in contrast, net rotation can occur under isotropic drag. We first demonstrate this result formally by considering the consequences of the force- and torque-free conditions on swimming bodies and we then illustrate it with two examples (a simple swimmers made of three rods and a model bacterium with two helical flagellar filaments). Our results highlight the different role of hydrodynamic forces in generating translational versus rotational propulsion.

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Since the 1950s [1,2], a continuous dialogue between theory and experiments has allowed us to unravel the fundamental physics of microorganism locomotion [3], and we can now predict how different organisms swim [4,5], how they move to favorable environments [6,7], and how they respond to boundaries [8,9]. The scientific community has also created its own series of synthetic microswimmers and attempted to optimize them [10–13].

Our understanding is made possible by our combined ability to (a) accurately measure the motion of microscale swimmers [4,7,9] and (b) theoretically describe the motion of the surrounding fluid through the incompressible Stokes equations [3,14]. The flow around the swimmer is obtained by enforcing that the fluid velocity on its surface is the same as the velocity of the swimmer itself (no-slip boundary condition) and the swimming kinematics are such that there is no net force and torque on the swimmer (free-swimming conditions). Since the Stokes equations are linear and time independent, net propulsion can only be created by a stroke kinematics, term nonreciprocal, which breaks the time symmetry of the system [15]. In the vast majority of cases, microorganisms generate nonreciprocal strokes by sending bending waves [16] or rotating [17] slender filaments termed flagella.

Such slender filaments are able to generate net propulsive forces due to their drag anisotropy at low Reynolds numbers (so-called drag-based thrust). Specifically, the drag per unit length acting on the slender filament is smaller for a translation along its centerline than for translation perpendicular to it. This is a fundamental property of small-scale fluid mechanics, which originates from the Green's function, $\mathbf{G}(\mathbf{r})$, for the incompressible Stokes equations due to a point force \mathbf{f} located at \mathbf{r}_0 [18],

$$\mathbf{G}(\mathbf{r}) = \frac{1}{8\pi\mu} \frac{\mathbf{I} + \hat{\mathbf{R}}\hat{\mathbf{R}}}{|\mathbf{R}|} \cdot \mathbf{f},$$
 (1)

where μ is the dynamic viscosity of the fluid, **I** is the identity tensor, and $\mathbf{R} = \mathbf{r} - \mathbf{r}_0$ is the vector pointing from the location of the point force to the point of interest ($\hat{\mathbf{R}}$ is a unit vector

in the same direction). Clearly, the flow resulting from the Green's function at any point in which \mathbf{R} is parallel to \mathbf{f} is twice as strong as the flow at a point with \mathbf{R} is perpendicular to \mathbf{f} for the same $|\mathbf{R}|$.

The resulting modeling approaches to describe the motion of slender filaments in viscous fluids therefore also display this feature of drag-anisotropy. The two commonly used theories are resistive-force (or local-drag) theory, which is analytical but only logarithmically correct [2,19,20], and slender-body theory, which has to be implemented numerically in general but is algebraically correct [12,21,22].

In a number of important situations, the anisotropy of the drag is less prominent. For example, some eukaryotic microorganisms have evolved hairs along their flagella called mastigonemes [23,24]. These change the drag characteristics of the filament by making the drag parallel to the filament similar to, or smaller than, the drag perpendicular to the filament, allowing these swimmers to swim "backwards." When these hairs are at the correct length and density, the drag on the filament therefore becomes isotropic, removing the anisotropic influence imparted by the Stokes Green's function. Similarly in non-Newtonian environments the drag changes in complex ways [25]. For example, the drag on rods in a shear thinning fluid was seen to decrease as the non-Newtonian nature of the fluid was increased [26]. This relative decrease was seen to be similar for motions parallel and perpendicular, though if the decrease was different for the two motions the system may again develop isotropic drag characteristics. Similarly, if the Stokes Green's function was made isotropic (i.e., proportional to the Laplacian Green's function), the drag on a filament would arise from a line distribution of these isotropic Green's functions and would be isotropic as a result.

However, drag anisotropy is the critical physical ingredient to allow motion of the geometric center of a swimmer, and without it a net translation is impossible [27]. Indeed, consider an inextensible [28] filament of length *L* described by the centerline location $\mathbf{r}(s,t)$ and deforming its shape with the instantaneous velocity $\mathbf{U}(s,t)$ in the laboratory frame of reference (Fig. 1). Under isotropic drag, the hydrodynamic force density along the filament, at arc length *s*, scales as $\mathbf{f}(s,t) \propto \mathbf{U}(s,t)$, so that the velocity of the mean filament

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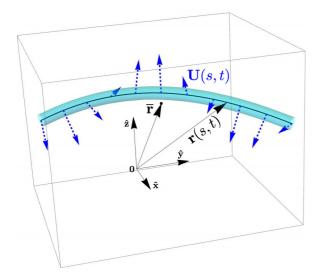


FIG. 1. Diagram showing the slender filament used to prove the necessity of drag anisotropy. The black line represents the filaments centerline, $\mathbf{r}(s,t)$, and the blue dotted arrows indicate the instantaneous velocity of the filament at arc length *s* and time *t*, $\mathbf{U}(s,t)$. The mean geometric position, $\mathbf{\bar{r}}(t)$, is also indicated for said filament.

position, $\overline{\mathbf{r}}(t) = (\int \mathbf{r}(s,t) \, ds)/L$, is given by

$$\frac{d\mathbf{\bar{r}}}{dt} = \frac{1}{L}\frac{d}{dt}\int_0^L \mathbf{r}(s,t)\,ds = \frac{1}{L}\int_0^L \mathbf{U}(s,t)\,ds$$
$$\propto \int_0^L \mathbf{f}(s,t)\,ds = \mathbf{0},\tag{2}$$

since the swimmer is force-free at all times. Allowing the body to be extensible can break this condition, prompting the creation of many popular theoretical models, like extensible filament swimming [28] and three sphere swimmers [29,30]. However, for many microswimmers, which are inextensible, drag anisotropy is a fundamental constraint on whether an organism can translate at low Reynolds number.

The argument shown in Eq. (2) applies to the swimmer's translation. It is unclear if a similar reasoning may be used to rule out a net rotation. In this paper we show that, in fact, drag anisotropy is not required to generate rotation. This is first shown formally by considering the force- and torque-free condition for the arbitrary deformation of a swimmer actuating slender appendages. We then illustrate the generation of rotation using model two swimmers: a lopsided paddle swimmer composed of three rods and a model bacterium with two flagellar filaments. Our work demonstrates that geometry alone can generate the conditions required to induce rotation and highlights the different role of hydrodynamic forces in generating translational versus rotational propulsion.

Using the notation above to describe the filament, we employ $\mathbf{U}_{def}(s,t)$ to denote the instantaneous zero-mean deformation of the body in the laboratory frame. We pick the instantaneous origin of the frame of reference to be the center of mass of swimmer and thus write $\mathbf{\bar{r}} = \mathbf{0}$. The velocity

along the shape of the swimmer is therefore written as

$$\mathbf{U}(s,t) = \mathbf{U}_{def}(s,t) + \mathbf{U}(t) + \mathbf{\Omega}(t) \times \mathbf{r}(s,t), \quad (3)$$

where $\mathbf{U}(t)$ and $\mathbf{\Omega}(t)$ are the instantaneous translation and rotational velocities. If motion occurs in a medium with isotropic drag, then Eq. (2) shows that $\mathbf{U}(t) = \mathbf{0}$ for all times. Denoting $\mathbf{f}(s,t) = \zeta \mathbf{U}(s,t)$ the isotropic relationship between the hydrodynamic force density acting on the fluid and the velocity of the swimmer centerline, we can write

$$\mathbf{f}(s,t) = \zeta \mathbf{U}_{def}(s,t) + \zeta \,\mathbf{\Omega}(t) \times \mathbf{r}(s,t), \tag{4}$$

where ζ is the isotropic drag per unit length. The form of ζ depends on the specific situation, whether that be a non-Newtonian fluid or a complex geometry and it is left general here to demonstrate this effect in any environment where isotropic drag on a "filament" is present. From the isotropic drag condition, the torque density, $\ell(s,t)$, is given by

$$\boldsymbol{\ell}(s,t) = \mathbf{r}(s,t) \times \mathbf{f}(s,t) + \gamma \, \boldsymbol{\widehat{\mathbf{tt}}} \cdot \boldsymbol{\Omega}(t), \tag{5}$$

where $\hat{\mathbf{t}} \equiv \hat{\mathbf{t}}(s,t)$ is the tangent vector along the body centerline. The last term in Eq. (5) accounts for the torque generated from local rotation about the centerline of the filament, with a rotational drag coefficient denoted γ (in both illustrative examples below we chose $\gamma = 0$, but it has been left here for completeness). We thus get a torque density given by

$$\boldsymbol{\ell}(s,t) = \zeta \mathbf{r}(s,t) \times [\mathbf{U}_{def}(s,t) + \boldsymbol{\Omega}(t) \times \mathbf{r}(s,t)] + \gamma \hat{\mathbf{t}} \hat{\mathbf{t}} \cdot \boldsymbol{\Omega}(t).$$
(6)

The total force and torque on the body are then given by

$$\mathbf{0} = \int_{0}^{L} \mathbf{f}(s,t) \, ds = \zeta \, \mathbf{\Omega}(t) \times \int_{0}^{L} \mathbf{r}(s,t) \, ds, \tag{7}$$
$$\mathbf{0} = \int_{0}^{L} \boldsymbol{\ell}(s,t) \, ds = \zeta \int_{0}^{L} \mathbf{r}(s,t) \times \mathbf{U}_{\text{def}}(s,t) \, ds$$
$$+ \int_{0}^{L} (\zeta \, \mathbf{I} r^{2} - \zeta \, \mathbf{r} \mathbf{r} + \gamma \, \mathbf{\hat{t}} \mathbf{\hat{t}}) \, ds \cdot \mathbf{\Omega}(t), \tag{8}$$

where we have used the vector identity $\mathbf{a} \times (\mathbf{b} \times \mathbf{c}) = \mathbf{b}(\mathbf{a} \cdot \mathbf{c}) - \mathbf{c}(\mathbf{a} \cdot \mathbf{b})$. The zeros on the left-hand side of Eqs. (7) and (8) reflect the fact that the swimmer is force- and torque-free for all times. The force-free condition is automatically satisfied since $\mathbf{\bar{r}} = \mathbf{0}$. The torque-free condition, Eq. (8), leads to an explicit equation for the rotation rate of the swimmer as

$$\mathbf{R}(t) \cdot \mathbf{\Omega}(t) = -\zeta \int_0^L \mathbf{r}(s,t) \times \mathbf{U}_{def}(s,t) \, ds, \qquad (9)$$

where the resistance tensor, $\mathbf{R}(t)$, is instantaneously given by

$$\mathbf{R}(t) = \int_0^L (\zeta \mathbf{I} r^2 - \zeta \mathbf{r} \mathbf{r} + \gamma \mathbf{\hat{t}} \mathbf{\hat{t}}) \, ds. \tag{10}$$

Hence, provided the deformation generates a net torque on the right-hand side of Eq. (9), and since the resistance tensor **R** is always positive definite for any finite-length filament, then $\Omega(t) \neq 0$ without the need for drag anisotropy. Note that this derivation is independent of the reference frame as it involves instantaneous velocities (the laboratory frame can be chosen

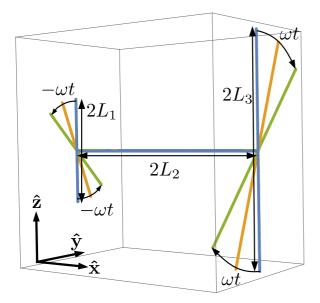


FIG. 2. Representation of the lopsided paddle swimmer with three arms of lengths $2L_1$, $2L_2$, and $2L_3$. The different colored arms show the swimmer configuration at different times with arrows indicating their rotation. The frame of reference (x, y, z) rotate with the swimmer so that the x axis is instantaneously aligned with rod #2.

to match the swimming frame at the time t, without loss of generality).

Physically, anisotropy is necessary to create any motion. In the case of linear velocity this anisotropy must come from the drag, as demonstrated by Eq. (2). However, in the case of rotation, the configuration of the centerline, $\mathbf{r}(s,t)$, can generate the required anisotropy, as shown by the nonisotropic resistance tensor **R**, even in the presence of isotropic drag. This instantaneous rotation can then generate a net rotation over a period if the deformation undergoes a nonreciprocal stroke, as per the scallop theorem [15].

To provide further intuition, we illustrate this result on two simple examples exhibiting nonzero angular velocities. Note that many more examples can be created from existing calculations that use resistive-force theory by formally setting the drag to be isotropic [4,31].

First we consider an elementary lopsided paddle swimmer as shown in Fig. 2. This swimmer consists of three straight rods of lengths $2L_1$, $2L_2$, and $2L_3$. The first and third rods are perpendicular to the second rod and both positioned at opposite ends of the second. The first and third rod then rotate around the axis of the second rod in opposing directions with period $T = 2\pi/\omega$. The centerline of this swimmer is described for all times and in the (x, y, z) frame rotating with it (see Fig. 2) as

$$\mathbf{r} + \bar{\mathbf{r}} = \begin{cases} \{-L_2, -s_1 \sin \omega t, s_1 \cos \omega t\} & -L_1 < s_1 < L_1, \\ \{s_2, 0, 0\} & -L_2 < s_2 < L_2, \\ \{L_2, s_3 \sin \omega t, s_3 \cos \omega t\} & -L_3 < s_3 < L_3, \end{cases}$$
(11)

where s_i describes the configuration of rod *i*. The origin of the reference frame is located at the center of the swimmer, $\mathbf{\bar{r}}$,

which is found by

$$2(L_1 + L_2 + L_3)\overline{\mathbf{r}} = \int (\mathbf{r} + \overline{\mathbf{r}}) ds$$

= $\int_{-L_1}^{L_1} \{-L_2, -s_1 \sin \omega t, s_1 \cos \omega t\} ds_1$
+ $\int_{-L_2}^{L_2} \{s_2, 0, 0\} ds_2$
+ $\int_{-L_3}^{L_3} \{L_2, s_3 \sin \omega t, s_3 \cos \omega t\} ds_3$
= $\{2L_2(L_3 - L_1), 0, 0\}.$ (12)

Hence, the center of the swimmer is a point on the second rod closer to the longest rotating rod. The deformation velocity for this swimmer is given by

$$\mathbf{U}_{def} = \omega \begin{cases} \{0, -s_1 \cos \omega t, -s_1 \sin \omega t\} & -L_1 < s_1 < L_1, \\ \{0, 0, 0\} & -L_2 < s_2 < L_2, \\ \{0, s_3 \cos \omega t, -s_3 \sin \omega t\} & -L_3 < s_3 < L_3, \end{cases}$$
(13)

and it generates an instantaneous net force and torque on the fluid of magnitudes

$$\mathbf{F} = \zeta \int \mathbf{U}_{\text{def}} \, ds = \{0, 0, 0\},\tag{14}$$

$$\mathbf{L} = \zeta \int \mathbf{r} \times \mathbf{U}_{\text{def}} \, ds = \left\{ -\frac{2}{3} \left(L_3^3 - L_1^3 \right) \zeta \, \omega, 0, 0 \right\}, \quad (15)$$

where the integrals are taken over all three rods as in Eq. (12). Assuming for simplicity that $\gamma = 0$, then the resistance matrix from Eq. (10) is instantaneously

$$\mathbf{R}(t) = \zeta \begin{pmatrix} \frac{2}{3} \left(L_1^3 + L_3^3 \right) & 0 & 0\\ 0 & \frac{2}{3} A(t) & B(t)\\ 0 & B(t) & \frac{2}{3} C(t) \end{pmatrix}, \quad (16)$$

where the coefficients A, B, and C are given by

$$A(t) = \frac{L_2^2 [4L_1(L_2 + 3L_3) + L_2(L_2 + 4L_3)]}{L_1 + L_2 + L_3} + (L_1^3 + L_3^3) \cos^2 \omega t, \qquad (17)$$

$$B(t) = -\frac{1}{3} \left(L_3^3 - L_1^3 \right) \sin 2\omega t, \qquad (18)$$

$$C(t) = \frac{L_2^2 [4L_1(L_2 + 3L_3) + L_2(L_2 + 4L_3)]}{L_1 + L_2 + L_3} + (L_1^3 + L_3^3) \sin^2 \omega t.$$
(19)

Inverting \mathbf{R} , we obtain that the instantaneous rotation rate of the lopsided paddle swimmer is constant and given by

$$\mathbf{\Omega} = \mathbf{R}(t)^{-1}\mathbf{L} = \left\{-\frac{L_3^3 - L_1^3}{L_3^3 + L_1^3}\omega, 0, 0\right\},\tag{20}$$

which has a nonzero angular velocity provided L_1 is not equal to L_3 . Clearly, if both rods have finite sizes, then the rotation rate of the whole swimmer is not equal to minus the rotation

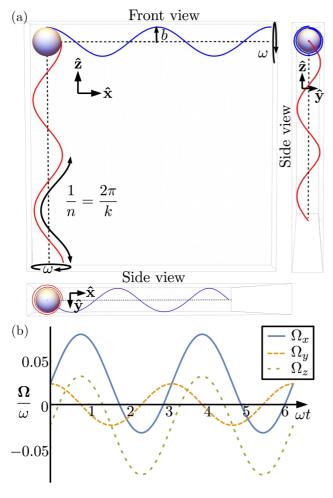


FIG. 3. Rotation of a model bacterium under isotropic drag. (a) Geometry of the swimmer composed of two identical helices of perpendicular axis, amplitude *b*, and wave number *k*, and rotate with frequency ω . The cell body is small compared to the helices and can thus be ignored. (b) Components of the angular velocity of the swimmer in the swimmer frame, Ω (scaled by ω), as a function of dimensionless time. For all figures b = 0.05 and $k = 4\pi$.

rates of each rod and indicates that a net rotation of the whole swimmer body can be induced purely from geometry.

As a second example, we consider a model for a bacterium with two flagellar filaments, as illustrated in Fig. 3(a). The swimmer is composed of two identical rigid helices attached at one end to a cell body and oriented with their helix axes perpendicular to each other. Each helix rotates around its axis with period $T = 2\pi/\omega$. For simplicity we ignore hydrodynamically the presence of the cell body, which is correct in the limit where the helical flagella are much longer than the body. In the frame of reference attached to the swimmer [see Fig. 3(a)], the location of each helix is given by

$$\mathbf{r}_1 + \overline{\mathbf{r}} = \{\alpha s, b \sin(kx - \omega t), b \cos(kx - \omega t)\}, \qquad (21)$$

$$\mathbf{r}_2 + \bar{\mathbf{r}} = \{b\cos(kx - \omega t), b\sin(kx - \omega t), -\alpha s\}, \quad (22)$$

where \mathbf{r}_1 and \mathbf{r}_2 denote, respectively, the centerlines of the first and second helices, 0 < s < 1, $k = 2n\pi$ is the wave number of the helix for a positive integer *n*, *b* its amplitude, and α is the cosine of the helix angle which satisfies $\alpha^2 + b^2k^2 = 1$ for inextensible helices. This swimmer rotates about the center $\overline{\mathbf{r}}$ defined as

$$\overline{\mathbf{r}} = \frac{1}{2} \int_0^1 (\mathbf{r}_1 + \overline{\mathbf{r}}) \, ds + \frac{1}{2} \int_0^1 (\mathbf{r}_2 + \overline{\mathbf{r}}) \, ds$$
$$= \frac{\alpha}{4} \{1, 0, -1\}, \tag{23}$$

which is the origin for both \mathbf{r}_1 and \mathbf{r}_2 . The deformation velocity for for each helix, $\mathbf{U}_{def,i}$, is then obtained by computing $\partial_t \mathbf{r}_i$ (i = 1,2). We obtain that the rotation of the helices around their axes generates net forces and torques on the body of magnitude

$$\mathbf{F}(t) = \zeta \int \mathbf{U}_{\text{def}} \, ds = \{0, 0, 0\},\tag{24}$$

$$\mathbf{L}(t) = \zeta \int \mathbf{r} \times \mathbf{U}_{\text{def}} \, ds \tag{25}$$

$$= \frac{b\zeta\omega}{2n\pi} \{2bn\pi + \alpha \sin \omega t, \alpha \cos \omega t, \alpha \sin \omega t - 2bn\pi\}.$$

Assuming $\gamma = 0$ for simplicity, the resistance matrix relating torque and rotation for this bacterial configuration can be computed exactly and we obtain

$$\mathbf{R}(t) = \zeta \begin{pmatrix} \frac{36b^2 + 5\alpha^2}{24} & \frac{b\alpha\cos\omega t}{2n\pi} & -\frac{\alpha^2}{8} \\ \frac{b\alpha\cos\omega t}{2n\pi} & b^2 + \frac{5}{12}\alpha^2 & -\frac{b\alpha\cos\omega t}{2n\pi} \\ -\frac{\alpha^2}{8} & -\frac{b\alpha\cos\omega t}{2n\pi} & \frac{36b^2 + 5\alpha^2}{24} \end{pmatrix},$$
(26)

and the torque-free condition then gives the swimmer an instantaneous angular velocity of

$$\mathbf{\Omega}(t) = -6b\omega \{A_2(t) + B_2(t), C_2(t), A_2(t) - B_2(t)\}, \quad (27)$$

where

$$A_2(t) = \frac{\alpha \sin \omega t}{n\pi (18b^2 + \alpha^2)},$$
(28)

$$B_2(t) = \frac{2bn^2\pi^2(12b^2 + 5\alpha^2) - 6b\alpha^2\cos^2\omega t}{n^2\pi^2(108b^4 + 69b^2\alpha^2 + 10\alpha^4) - 36b^2\alpha^2\cos^2\omega t},$$
(29)

$$C_2(t) = \frac{2n\pi\alpha(3b^2 + 2\alpha^2)\cos\omega t}{n^2\pi^2(108b^4 + 69b^2\alpha^2 + 10\alpha^4) - 36b^2\alpha^2\cos^2\omega t}.$$
(30)

We plot in Fig. 3(b) all the components of Ω , nondimensionalized by ω , for the values b = 0.05 and $k = 4\pi$ (i.e., n = 2). We see that the instantaneous rotational velocity is nonzero for all body directions and oscillates sinusoidally around a mean value of

$$\frac{\omega}{2\pi} \int_0^{2\pi/\omega} \mathbf{\Omega} \, dt = \omega \bigg[(1 - \sqrt{W}) + \frac{6r_h^2}{9r_h^2 + 2\alpha^2} \sqrt{W} \bigg] \{-1, 0, 1\},$$
(31)

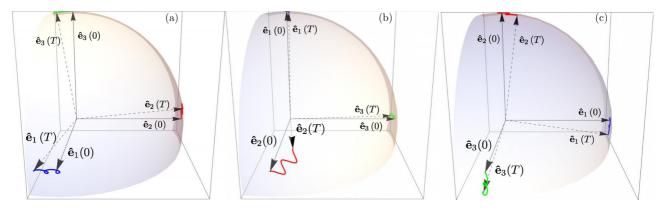


FIG. 4. Trajectories of the body vectors $(\hat{\mathbf{e}}_1(t), \hat{\mathbf{e}}_2(t), \hat{\mathbf{e}}_3(t))$ on the surface of a unit sphere $(r_h = 0.05, k = 4\pi)$. (a) The trajectory of $\hat{\mathbf{e}}_1(t)$ (blue); (b) The trajectory of $\hat{\mathbf{e}}_2(t)$ (red); (c) The trajectory of $\hat{\mathbf{e}}_3(t)$ (green). The paths over the sphere represent the motion with the initial (solid) and final (dashed) vectors clearly labeled. In the above $T = 2\pi/\omega$.

where

$$W = \frac{108r_h^2 + \alpha^2 (69 - 108r_h^2) - \alpha^4 (69 - 10n^2 \pi^2)}{108r_h^2 + \alpha^2 (69 - 72r_h^2) - \alpha^4 (69 - 10n^2 \pi^2)}$$
(32)

The angular displacement experienced in the laboratory frame can be found from

$$\frac{d\hat{\mathbf{e}}_i}{dt} = \mathbf{\Omega}(t) \times \hat{\mathbf{e}}_i, \qquad (33)$$

where $\hat{\mathbf{e}}_i$ is the body frame vector i = 1, 2, 3. We take $\hat{\mathbf{e}}_1(t)$ to be the body vector aligned with the helix axis of \mathbf{r}_1 , $\hat{\mathbf{e}}_3(t)$ to be the body vector aligned with the helix axis of \mathbf{r}_2 , and $\hat{\mathbf{e}}_2(t)$ to be the body vector perpendicular to the helix axes. The above equations were solved numerically for a bacterial swimmer with $r_h = 0.05$ and $k = 4\pi$ [32]. In this configuration, the body vectors, after one period of rotation, become

$$\hat{\mathbf{e}}_1\left(T = \frac{2\pi}{\omega}\right) = \{0.989. - 0.149, -0.003\},$$
 (34)

$$\hat{\mathbf{e}}_2\left(T = \frac{2\pi}{\omega}\right) = \{0.148, 0.9778, 0.149\},$$
 (35)

$$\hat{\mathbf{e}}_{3}\left(T=\frac{2\pi}{\omega}\right) = \{-0.020, -0.148, 0.989\},$$
 (36)

where we have assumed $\hat{\mathbf{e}}_1(0) = \{1,0,0\}, \hat{\mathbf{e}}_2(0) = \{0,1,0\}$, and $\hat{\mathbf{e}}_3(0) = \{0,0,1\}$. These vectors are written in terms of the *x*, *y*, and *z* coordinates of the laboratory frame. Figure 4 plots

the trajectories of these vectors on the surface of a unit sphere. Hence, under isotropic drag, this model bacterium undergoes a nontrivial net rotation in the laboratory frame due solely to the anisotropy of its shape.

Anisotropy in their linear drag is a requirement for microorganisms using filamentous appendages to undergo net translation through viscous fluids. By extension one may think that such drag anisotropy is also required to generate rotation. In this paper we showed that in fact rotation was possible in a system with isotropic drag. This result was evident when considering the force- and torque-free conditions directly and allowed us to demonstrate that in such a fluid the rotation is physically generated by the anisotropy in the shape of the swimmer (specifically, the centerline of the slender filaments it actuates). We derived this rotation for an arbitrary slender body and then illustrated it on two model swimmers, a lopsided paddle swimmer and a multiflagellated model bacterium, which both exhibit nonzero rotation within an isotropic-drag medium. Other examples may also be created by making the drag isotropic in existing resistive force theory studies [4,31]. Our results highlight the different role of hydrodynamic forces in generating translational versus rotational propulsion and may change our understanding of the physical requirements for rotational motion in complex environments.

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